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Ubiquitin Ligase HUWE1 Regulates Axon Branching through the Wnt/β-Catenin Pathway in a Drosophila Model for Intellectual Disability

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Abstract

We recently reported that duplication of the E3 ubiquitin ligase HUWE1 results in intellectual disability (ID) in male patients. However, the underlying molecular mechanism remains unknown. We used Drosophila melanogaster as a model to investigate the effect of increased HUWE1 levels on the developing nervous system. Similar to the observed levels in patients we overexpressed the HUWE1 mRNA about 2-fold in the fly. The development of the mushroom body and neuromuscular junctions were not altered, and basal neurotransmission was unaffected. These data are in agreement with normal learning and memory in the courtship conditioning paradigm. However, a disturbed branching phenotype at the axon terminals of the dorsal cluster neurons (DCN) was detected. Interestingly, overexpression of HUWE1 was found to decrease the protein levels of dishevelled (dsh) by 50%. As dsh as well as Fz2 mutant flies showed the same disturbed DCN branching phenotype, and the constitutive active homolog of β-catenin, armadillo, could partially rescue this phenotype, our data strongly suggest that increased dosage of HUWE1 compromises the Wnt/β-catenin pathway possibly by enhancing the degradation of dsh.

Introducution

Intellectual disability (ID) refers to cognitive impairment and affects ~2% of the population in developed countries. ID patients lack the necessary mental capabilities and adaptive skills required to live independently and rely on family members and other caretakers for help in daily life. ID therefore constitutes an important medical and socio-economic problem. Although numerous ID genes have been discovered during the last two decades, for many of them the molecular mechanism via which they contribute to the ID phenotype remains unknown. Particularly unexplored are mechanisms that underlie copy number gains in ID, increasing gene dosage, as most of the ID genes that have been investigated were studied in knockout animal models.

We recently identified non-recurrent but overlapping microduplications at Xp11.22 in 12 unrelated families with mild to moderate non-syndromic ID [1,2]. The only coding gene present in the smallest region of overlap of these microduplications is HUWE1. The region also contains two miRNAs, but the identification of a partial HUWE1 duplication harboring both miRNAs in healthy individuals strongly suggest that these do not contribute to ID phenotype [1]. These findings thus indicate that a modest increase in expression of HUWE1, 1.6- to 2.0-fold as observed in patients, is sufficient to cause non-syndromic cognitive impairment. Since all patients of the
reported 12 families show only mild to moderate ID irrespective of the size of the duplication, HUWE1 appears to be the main gene underlying the cognitive deficits. Further evidence for a role of HUWE1 in ID comes from missense mutations in this gene that we detected in 3 ID families [2]. The patients of these families presented with a more severe form of ID with additional syndromic features. However, complete loss-of-function of mouse Huwe1 in the brain leads to neonatal lethality [3,4]. Deletion of HUWE1 in the human population has never been reported.

The HECT, UBA and WWE domain containing 1 (HUWE1) gene encodes a ~500 kDa E3 ubiquitin ligase that targets its substrates for proteasomal degradation via poly-ubiquitination. Conditional Huwe1 knockout studies in the mouse have proven the importance of this protein for neuronal development via its role in the regulation of Mycn, which is essential for the initiation of cell cycle exit and start of neuronal differentiation in the brain [3,4]. However, no studies have yet been performed on the effects of increased HUWE1 expression in vivo. We used Drosophila melanogaster to investigate neuronal phenotypes and their underlying mechanisms/pathways. In the last decade, the fruitfly has emerged as a valuable model system for the study of ID [5,6]. In addition, it is especially suited for the investigation of increased expression in a tissuespecific manner because of the easy-to-use UAS-Gal4 system [7]. We generated HUWE1 transgenic flies, which did not show severe neurological or behavioral alterations. However, at single axon resolution, increased HUWE1 levels were found to disturb terminal branching of the dorsal cluster neurons (DCN), most likely by disturbing the canonical Wnt/β-catenin pathway, a mechanism that has not been significantly investigated in association with ID.

Materials and Methods

Fly strains and generation of transgenic lines

Fly stocks were cultured on standard fly food and crosses were set up according to standard procedures. All experiments were performed in temperature-controlled incubators at 25°C (or 28°C when mentioned). To create the UAS-HUWE1 line, we cloned the human HUWE1 cDNA (kindly obtained from V. Kalscheuer, Berlin) in the pUAST-attB vector followed by injection of this construct in Drosophila embryos by GenetiVision (www.genetivision.com). The construct was integrated in the fly genome via PhiC31-mediated site-specific integration at either insertion site VK37 at chromosome 2 or VK31 at chromosome 3 [8]. The transgenic flies will be referred to as UAS-HUWE1VK37 for insertion on 2L and UAS-HUWE1VK31 for insertion on 3L. Flies containing the AttP locus without a construct inserted were used as controls: controlVK37 at chromosome 2 or VK31 at chromosome 3 [8]. The transgenic flies will be referred to as UAS-HUWE1VK37 for insertion on 2L and UAS-HUWE1VK31 for insertion on 3L. Flies containing the AttP locus without a construct inserted were used as controls: controlVK31 is y w;P{Bac[y+attP-3B]VK00031}, and controlVK37 is y w;P{Bac[y+attP-3B]VK00037}. Other stocks used in this study are: nSyb-Gal4, elav-Gal4, Eyeless-Gal4; 201Y-Gal4, UAS-DenMark, UAS-mCD8-GFP, 247-Gal4, UAS-dfz2GPI; UAS-mCD8-GFP, ptdGal4-14a, UAS-LacZ, Dsh-Dsh-GFP, CyO, Dsh-Dsh-GFP, UAS-HUWE1VK31, Dsh-Dsh-GFP, UAS-armacr and Canton S10. The KK101525 Dsh RNAi line was obtained from the Vienna Drosophila RNAi Center (VDRC; Vienna, Austria) [9]. Additional information on Drosophila stocks and genes can be found in Flybase (http://flybase.org/).

Confirming the localization of UAS-HUWE1 constructs by PCR

The correct integration of the UAS-HUWE1 construct in the fly genome was confirmed by standard PCR with GoTaq polymerase (Promega) using primer HUWE1_ex24 localized in HUWE1 in combination with either primer Dros.VK37 on chromosome 2L for UAS-HUWE1VK37, or primer Dros.VK31 on 3L for UAS-HUWE1VK31. Primer sequences are available in Table S1.

Expression analysis by RT-qPCR

Total RNA was isolated from fly heads via TRizol (Invitrogen) – chloroform extraction, and cDNA was transcribed with the Quantitect Reverse transcription Kit (Qiagen) on 1 µg RNA. qPCR was performed on the LightCycler 480 system (Roche), and each 15 µl reaction well contained 5 µl of a 1/60 dilution of cDNA, 0.5 µM of each primer and 1x SYBRgreen qPCR Master Mix (Roche). All samples were run in duplicate, and specificity of the PCR products was checked by a melt-curve analysis. The data were analyzed with the LC480 software followed by analysis in Microsoft Excel via the comparative ddCt method (Applied Biosystems, Foster City, CA), with normalization of the expression levels to the housekeeping gene rp49. In order to compare expression of the human HUWE1 and the Drosophila homolog CG8184, we cloned the qPCR amplicons of HUWE1_ex73-74_for/rev and CG8184_for/rev together in the pGEM-T Easy vector (Promega). qPCR on this construct allowed us to compare the primer efficiency of both primer pairs and thus the accurate comparison of the expression levels of both genes. Primer sequences are available in Table S1.

Immunohistochemistry on whole mount fly brain

Adult brains were dissected in 1x PBS and fixed with 4% formaldehyde in PBT (1x PBS + 0.3% Triton X-100) for 15 min. After two washes with PBT, the fixed brains were blocked for one hour in PAXDG buffer (PBT, 5% normal goat serum, 1% bovine serum albumin, 0.1% deoxycholate, 1% Triton X-100) and then incubated overnight at 4°C with primary antibodies diluted in 1x PAXDG. The following primary antibodies were used: mouse anti-GFP mAb 3E6 (Invitrogen cat. no. A11120) 1:1000, Rabbit polyclonal anti-DsRed (Clontech cat. no. 632496) 1:1000, mouse FabII 1D4 (DSHB) 1:50, and rabbit anti-GFP (Invitrogen cat. no. A11122) 1:500. This incubation was followed by 5 wash steps of 10 min each in PBT and a final incubation of 3 h at RT with the appropriate fluorescent secondary antibodies (Alexa 488, 555 or 647, Molecular probes; 1:500). After 6 washes of 10 min in PBT the samples were mounted in Vectashield Mounting Medium. Images of adult brains were acquired on a Leica TCS SP5 II confocal microscope system (Leica Microsystems) equipped with 458, 476, 488, 514, 543 and 633 nm lasers, and processed using ImageJ [10]. To quantify the branch number of DCN axons, we used the Simple Neurite Tracer plug-in in Fiji [11].
**Protein quantification via Western Blotting**

For determination of Dsh levels, we lysed four adult brains of each genotype in 10 µl 2x sample buffer (2% SDS, 10 mM Tris pH 6.8, 1 mM EDTA, 10% glycerol, 0.05% Bromophenolblue). The samples were boiled for 10 min at 95°C, after which 1 µl of DTT was added. The samples were run on a 4-12% NuPAGE Bis-Tris precast polyacrylamide gel (Invitrogen) in MOPS buffer and electrophoretically transferred to Hybond-C extra nitrocellulose membrane (GE Healthcare). After blocking for 1 h with 5% nonfat milk, the membrane was incubated overnight at 4°C with mouse anti-GFP (Roche) 1:1000 and mouse anti-actin JLA20 (DSHB) 1:100 as a loading control. After washing with PBT, the blot was probed with sheep anti-mouse IgG secondary antibody conjugated to HRP at 1:1000 dilution (GE Healthcare) for 1 h at RT. The bands were visualized with ECL western blotting detection reagents (GE healthcare) and digitally imaged with the Fujifilm LAS-300 Mini system (Life Science Systems). The experiment was repeated 4 times with flies kept at 25°C and 2 times with flies kept at 28°C. Quantitation was performed with ImageJ [10].

**Courtship conditioning assay**

Double transgenic ;UAS-HUWE1VK37,UAS-HUWE1VK31 flies crossed to the mushroom body-specific 247-Gal4 driver were tested for learning and memory using the courtship conditioning assay as previously described [12]. Briefly, virgin males raised at 28°C were placed separately in food chambers together with a single mated female for a training period of 5 h. After training, all males were recovered and tested immediately to assess learning, or after 1 h to test short-term memory. All tests were carried out by pairing each male with a fresh mated female in a 1-cm courtship chamber for 10 min. The tests were videotaped and all assays were scored with customized tracking software from Actual Analytics (Edinburgh, UK). The mean Courtship Index (CI, the percentage of time spent on courtship during a 10 min interval) of trained males and of socially naïve males was used to calculate the Learning Index (LI), which is defined as the percent reduction in mean courtship activity in trained males compared with naïve males; LI = (O(CI naïve)−O(CI trained))/O(CI naïve). No significant difference was found.

**Neuromuscular junction (NMJ) and electroretinogram (ERG) analysis**

3rd instar larval grown at 28°C were dissected in PBS and fixed for 30 min in 3.7% paraformaldehyde. The fixed files were incubated overnight at 4°C with mouse anti-brp (nc82, DSHB, 1:125 dilution). To visualize brp/nc82, secondary Alexa 488 goat-anti-mouse mAb was used (Invitrogen, 1:500 dilution). Subsequently, anti-disks large 1 (dlg1) (DSHB) pre-labeled with the Zenon Alexa Fluor 568 Mouse IgG1 labeling kit (Invitrogen) was applied. Acquired images were automatically processed and measured by an advanced in-house developed Fiji-based macro (Figure S1). The analysis was performed on NMJs of muscle 4 of at least 22 synaptic terminals.

**Results**

**Generation of a fly model with HUWE1 overexpression**

After cloning the cDNA of the human HUWE1 gene in the pUAST-attB vector, site-specific integration via PhiC31 integrase was used to incorporate the construct in a well-defined AttP site on either the 2rd or 3rd chromosome of the fly. HUWE1 expression can easily be regulated in a time- and tissue-specific manner by crossing the UAS-HUWE1 transgenic fly to a particular Gal4 driver line. The correct location of the UAS-HUWE1 construct in the fly genome was confirmed by PCR, and expression of human HUWE1 was checked by RT-qPCR with three primer pairs distributed over the entire length of the mRNA. Both transgenic lines expressed human HUWE1 from a single allele in the fly head at ~1.9 fold the expression level of the endogenous homolog CG8184 when crossed to the pan-neuronal nSyb-Gal4 driver line. Both VK37 and VK31 lines were also crossed to generate a fly carrying the UAS-HUWE1 construct on both 2rd and 3rd chromosome. Crossing this double transgenic line to the neuron-specific ;elav-Gal4;elav-Gal4 driver line at 28°C resulted in an expression level of ~1.4 times the expression of CG8184. In comparison, the same driver line combined with the UAS-HUWE1VK31 line resulted in a HUWE1 expression that reached only about half the expression levels of the fly homolog CG8184. In ID patients with Xp11.22 duplications, the HUWE1 mRNA levels in blood lymphocytes were increased 1.6- to 2.0-fold. Flies expressing the HUWE1 via the neuron-specific elav-Gal4 or nSyb-Gal4 drivers from two alleles were viable and showed no overt morphological defects, which corresponds to

**Statistical analysis**

Data analysis was performed with Excel and GraphPad Prism 5. To compare the number of DCN axon branches in UAS-HUWE1, Dsh RNAi, Dsh and Dsh lines with control flies, we used 1-way ANOVA followed by Dunnett’s multiple comparison test. To compare branch number of UAS-HUWE1VK31, arm RNAi, UAS-armGT and UAS-armACT;UAS-HUWE1VK31 lines with control flies and UAS-HUWE1VK31 with UAS-armACT;UAS-HUWE1VK31, we used 1-way ANOVA followed by Bonferroni’s multiple comparison test. ControlVK37 and controlVK31 flies were used as controls. For statistical analysis of dsh protein quantitation and NMJ experiments, we used the two-tailed unpaired Student’s t-test. In the courtship conditioning assay, non-parametric statistical comparison of HUWE1 overexpressing and control flies was performed using a custom SAS script (SAS Institute, Inc.) to perform bootstrapping as described [14]. Briefly, CI values were randomly sampled with replacement to generate 10,000 hypothetical LIs, which were used to determine the 95% confidence interval of the difference between L((control) and LI(knockdown)).
the nonsyndromic phenotype of the patients. To study a potential effect of increased HUWE1 dosage on brain development and function in the fly, we first analyzed the flies for general neurological defects.

Learning and memory in the Courtship conditioning paradigm

As overexpression of HUWE1 in humans was suggested to impair cognition to a level below an IQ of 70, we first subjected the transgenic flies to the courtship conditioning paradigm. The assay is based on the suppression of courtship behavior observed in male flies after they have been rejected by a pre-mated female [12]. Because the mushroom bodies (MB) are known to play an important role in this form of learning and memory and in cognition in general, we used the MB-specific 247-Gal4 line to drive HUWE1 expression from the double transgenic ;UAS-HUWE1\(^{VK37}\);UAS-HUWE1\(^{VK31}\) line. The HUWE1 line had been outcrossed for at least 7 generations before the assay was performed (Figure S2). However, defects in synaptic plasticity cannot be excluded.

Morphology of the mushroom body

To examine if overexpression of HUWE1 had an effect on the structure of the nervous system we then investigated the MB for morphological alterations. The MB is formed by ~2500 Kenyon cells in each brain hemisphere that can be divided in 3 classes of neurons: γ neurons, α'/β' neurons and α/β neurons. The γ neurons project a single horizontal axon, whereas the α'/β' and α/β neurons axons branch in two projections, one horizontal and one vertical. Expression of HUWE1 was driven by 201Y-Gal4, a line that expresses extensively in the γ neurons of the MB and also in a small subset of the α/β neurons [15]. These neurons were visualized by expression of UAS-driven membrane-associated MCD8-green fluorescent protein (GFP). No aberrations in the axon lobes were detected (Figure 1B’-B’). In addition, we also specifically labeled the calyx, which is formed by the dendrites of the Kenyon cells, with the somatodendritic marker DenMark [16]. Also this dendritic structure did not present obvious morphological abnormalities (Figure 1B’-B’). We confirmed the absence of morphological abnormalities in the axon lobes with the MB-specific 247-Gal4 line driving HUWE1 expression from the double transgenic line ;UAS-HUWE1\(^{VK37}\);UAS-HUWE1\(^{VK31}\) (Figure 1C-C’). The lobes were visualized with anti-Fasll staining but no significant differences compared to the controls were detected. Since the MB is a dense structure formed by a large number of neurons, it is possible that more subtle aberrations remained unnoticed.

Analysis of the neuromuscular junctions and basal neurotransmission

Next, we examined the larval neuromuscular junction (NMJ), which is the most accessible synapse in the fly [17]. The Drosophila NMJ shares important features with central excitatory synapses in the vertebrate brain, as the NMJ is a glutamatergic synapse with ionotropic glutamate receptors that are homologous to those of humans [18]. We used the pan-neuronal ;UAS-dicer2; elav-Gal4 line to drive HUWE1 expression from the double transgenic ;UAS-HUWE1\(^{VK37}\);UAS-HUWE1\(^{VK31}\) line and examined the NMJ of muscle 4 in wandering 3rd instar larvae. No morphological abnormalities were detected in the NMJ: the NMJ area, perimeter and length were not affected and there were no differences in the number of branches and branching points (Table S2). We also investigated the number of active zones, which are the presynaptic sites of neurotransmitter release. The number of active zones, visualized by anti-nc82 staining, was not affected by increased HUWE1 levels (Table S2).

Basal neurotransmission can be detected by recording an electroretinogram (ERG), which assesses if synaptic transmission between photoreceptors and their post synaptic targets in the lamina in response to a light flash occurs and is synchronized. An ERG shows the voltage difference between the retina and the rest of the body during a short (1 s) light pulse. Adult flies expressing HUWE1 in their entire nervous system by the nSyb-Gal4 driver did not present any abnormalities in the ERG profile, indicating that neurotransmission and neuronal connectivity are largely unaffected (Figure S2). However, defects in synaptic plasticity cannot be excluded.

Axon branching in the dorsal cluster neurons

As our analyses above did not reveal any major alterations upon moderate HUWE1 overexpression, we decided to investigate if defects at a more sensitive level might explain why increase in HUWE1 levels causes cognitive problems in ID patients. For this we made use of the dorsal cluster neurons (DCNs) as a model because subtle changes can be detected at single axon resolution. The DCN consist of a small cluster of neurons whose axons grow in a very stereotypical pattern allowing the detection of subtle defects in axon outgrowth and branching. HUWE1 was expressed specifically in these neurons by means of the atoGal4-14a driver line. This driver starts to be expressed in early third instar larvae, which is shortly before the DCNs begin to extend their axons towards the optic lobes, and continues to be active during metamorphosis and into adult life [19,20]. The neurons were visualized by expression of the membrane-associated mCD8-GFP.

In wild-type adult flies, 11-12 parallel axons cross the optic chiasm between lobula and medulla, after which they branch to form a stereotypical grid-like structure [20]. The number of axons crossing the optic chiasm is not affected in flies with increased HUWE1 levels, with an average of 11.3 axons innervating the medulla for UAS-HUWE1\(^{VK37}\) versus 11.1 in control\(^{VK37}\), and 11.7 axons in UAS-HUWE1\(^{VK31}\) versus 11.9 in control\(^{VK31}\). However, the branching pattern in the medulla was
disturbed giving rise to an increased number of branches at the 3rd branching point of the grid-like structure (Figure 2). The effect was seen with UAS-HUWE1VK37 as well as with UAS-HUWE1VK31, indicating that the phenotype is specifically due to increased levels of HUWE1 and does not occur as a result of position effects caused by the insertion of the construct in the fly genome. Moreover, overexpression of another seemingly dosage-sensitive ID gene GDI1 that we tested with the same DCN-specific driver did not affect axon branching (data not shown), pointing to specificity of the phenotype for increased HUWE1 levels.
Next, we investigated the pathway that could affect axon branching upon \textit{HUWE1} overexpression. \textit{Huwe1} has recently been demonstrated to ubiquitinate dishevelled (\textit{dsh} in flies, \textit{Dvl} in mammals), a major component of the Wnt/\beta-catenin pathway (V. Bryja, personal communication). To test whether increased \textit{HUWE1} expression in the fly brain, driven by the pan-neuronal \textit{nSyb-Gal4} driver, had an effect on \textit{dsh} levels, we expressed GFP-tagged \textit{dsh} under its own promoter, allowing us to visualize \textit{dsh-GFP} with an anti-GFP antibody. Western blot detected a 50\% reduction in \textit{Dsh-GFP} levels in \textit{dshdsh-GFP/+;UAS-HUWE1\textsubscript{VK31}/nSyb-Gal4} flies compared to the controls (Figure 3). These data thus suggest that the Wnt/\beta-catenin pathway is also involved in the fly model presented here. If so, lower levels of the \textit{Drosophila} \textit{dsh} should result in a similar increased branching phenotype. Indeed, flies targeting \textit{dsh} expression via RNA interference (RNAi) presented a significant increase in branch number at the 3\textsuperscript{rd} branching point of the DCNs, strikingly similar to \textit{HUWE1} overexpressing conditions (Figure 4A). The same phenotype was observed in flies heterozygous for \textit{dsh\textsuperscript{6}}, a null allele of \textit{dsh} (Figure 4B). Homozygous null mutants could not be tested due to embryonic lethality. We also investigated the effect of the \textit{dsh\textsuperscript{1}} mutant on DCN axon branching. This mutant is deficient only in the activity of its DEP domain (Dishevelled, Egl-10, Pleckstrin), which is important for the activation of the non-canonical JNK signaling pathway by regulation of Rho family GTPase proteins. In contrast, it leaves the DIX domain (Dishevelled and Axin) intact, which is specifically required for the activation of the canonical \beta-catenin pathway [21,22]. DCN axon branching at the 3\textsuperscript{rd} branching point was not affected in \textit{dsh\textsuperscript{1}} mutant males.
indicating that activation of the non-canonical pathway via the DEP domain does not play a role in this branching process. A second line of evidence for the involvement of the Wnt/β-catenin pathway was observed upon expression of a dominant-negative mutant form of the Wnt-receptor frizzled2 (fz2), which resulted in a similar disturbed DCN branching phenotype as observed for HUWE1 overexpression (Figure 4D). As illustrated in Figure 4G activation of the frizzled (fz) receptor upon binding of its Wnt ligand will lead to inhibition of the β-catenin destruction complex via activation of dsh. β-catenin can then activate its target genes, which could in this case either stop the branching process or initiate pruning of excessive branches. Expression of the dominant-negative fz2 prevents this activation and thus causes the observed increase in axon branch number. Finally, the involvement of the Wnt/β-catenin pathway was investigated via the Drosophila homolog of β-catenin, armadillo (arm). We were able to partially rescue the increased branching phenotype by combining HUWE1 overexpression with expression of a constitutively active mutant of arm (Figure 4E). This mutant can no longer be inactivated by the β-catenin destruction complex, and as a consequence it is not affected by the HUWE1-driven breakdown of dsh. These data are in agreement with the expression of a constitutively active arm, which on its own resulted in a reduced branching phenotype (Figure 4F). Quantification of branching relative to the controls is shown in Figures 4H-I, which also shows that RNAi for arm does not affect branching. In conclusion, we provide evidence that overexpression of HUWE1 lead to increased breakdown of dsh, causing the β-catenin destruction complex to remain more active and thus reduce the activation of β-catenin-dependent genes (Figure 4G).

Figure 3

Figure 3. HUWE1 overexpression reduces dsh levels. (A) HUWE1 expression was driven by the pan-neuronal nSyb-Gal4 driver, and whole-brain lysates were subjected to SDS-PAGE. Dsh-GFP was visualized with an anti-GFP antibody, Actin was used as loading control. Lane 1: dsh<dsh-GFP/+;nSyb-Gal4/+; Lane 2: dsh<dsh-GFP/+;UAS-HUWE1VK31/nSyb-Gal4. (B) Quantitation of dsh-GFP levels, normalized to actin. The graph represents the average of 4 biological repeats with SEM. Dsh-GFP levels are significantly reduced in flies with HUWE1 overexpression (** p<0.01, Student’s t-test). Control = dsh<dsh-GFP/+;nSyb-Gal4/+; HUWE1 = dsh<dsh-GFP/+;UAS-HUWE1VK31/nSyb-Gal4.

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Figure 4. The Wnt/β-catenin pathway is involved in the disturbed DCN branching. (A,B) Reduced dsh levels in w;UAS-dshRNAi/UAS-mCD8-GFP;atoGal4-14a,UAS-LacZ/+ and heterozygous null mutant dsh^{6}+/;UAS-mCD8-GFP/+;atoGal4-14a,UAS-LacZ/+ animals also led to an increased axon branching at the 3rd branching point. (C) DCN axon branching is normal in dsh^{1};UAS-mCD8-GFP/+;atoGal4-14a,UAS-LacZ/+ males, which are only mutant in the DEP domain responsible for activation of the non-canonical pathway. (D) DCN axon branching is equally affected in dominant negative Fz2;UAS-dn-Fz2/UAS-mCD8-GFP;atoGal4-14a,UAS-LacZ/+ flies. (E) Combined expression of HUWE1 and Arm^{AC} in w;UAS-Arm^{AC}/UAS-mCD8-GFP;UAS-HUWE1^{VK31}/atoGal4-14a,UAS-LacZ flies partially rescues the branching phenotype, although the number of branches is not completely reverted to wild-type levels. (F) Expression of a constitutively active Arm mutant in w;UAS-Arm^{AC}/UAS-mCD8-GFP;atoGal4-14a,UAS-LacZ/+ animals results in a reduced branching phenotype. (G) Model showing the association of HUWE1 with the Wnt/β-catenin pathway and its effect on axon branching and/or pruning, as evidenced by our data. (H,I) Quantitation of the axon branching levels at the 3rd branching point of the DCNs in the medulla. We evaluated 20-25 neurons from at least 5 different brains per genotype. Error bars represent standard error of the mean (SEM) (** p<0.01, *** p<0.001).

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Discussion

In this study we developed a *Drosophila melanogaster* model to investigate the effect of increased HUWE1 expression on the developing nervous system. Quantitation of HUWE1 levels indicated that expression in the fly brain was increased in a range similar to what is seen in the patients, who presented 1.6- to 2-fold increase levels compared to controls [2]. For instance, the pan-neuronal elav-Gal4;elav-Gal4 driver line resulted in a HUWE1 expression that reached about half the levels of the fly homolog CG8184. Assuming that human HUWE1 and *Drosophila* CG8184 perform the same functions, the transgenic fly has a combined HUWE1-CG8184 level that is 1.5 fold higher compared to that of the endogenous CG8184 expressed in control flies. This increased dosage is in the same range as the 1.5- to 1.8-fold increase described for dap160, synj and nla in a fruitfly model for Down syndrome. Even this as well in flies with a pan-neuronal increased expression of HUWE1 overexpressing biological processes [24], it has been associated with terminal neurons that are part of the courtship circuit to cause a phenotype. Furthermore, no broad deregulation was noticed in the MB. This finding could be in accordance with the fact that MRI brain imaging and CT scans of patients with HUWE1 duplications did not reveal gross structural aberrations [2]. Morphology of the NMJs and neurotransmission, measured as ERGs, were unaffected in flies with a pan-neuronal increased expression of HUWE1. These data are in agreement with the lack of additional clinical features in the patients as HUWE1 is a ubiquitously expressed gene.

By looking for more subtle alterations however, we detected a significant increase in branch numbers at the axon terminals of the dorsal cluster neurons. The DCNs are a group of ~40 neurons that are part of the *Drosophila* visual system. Their cell bodies are located in the dorso-lateral region of the central brain, from where they project their axons to the contralateral side of the brain to innervate the optic lobe [15,16]. Our data suggest that this altered branching phenotype was caused via a negative regulatory effect of HUWE1 on dsh, a key component of the Wnt signaling pathway that transmits Wnt signals from the fz receptors to downstream effectors. Though Wnt signaling plays an important role in a wide range of biological processes [24], it has been associated with terminal branching of neurons too. Wnt5a and Wnt3 have previously been implicated to regulate the axon branching in the mouse sympathetic and spinal sensory dorsal root ganglia neurons, respectively [25,26]. In *Drosophila*, axon branching of the MB neurons was shown to be regulated by Wnt signaling [27]. Altered Wnt signaling has also been linked to synaptic plasticity in the mouse [28] and to other neurological disorders such as Alzheimer’s disease [29], Williams syndrome [30] and schizophrenia [31,32], as well as neurodevelopmental abnormalities including neural tube defects and agenesis of the corpus callosum [24]. Interestingly, ubiquitination has been shown to play an important role in the regulation of Wnt signaling by affecting many different steps in the pathway, as reviewed by Tauriello and Maurice [33]. In fact, the E3 ubiquitin ligase Nedd4 was demonstrated to promote axon branching in Xenopus retinal ganglion cells [34]. Recently, HUWE1 was identified as a conserved negative regulator of the Wnt/β-catenin pathway acting at the level of Dvl (V. Bryja, personal communication) and conditional knock down of different key components of the Wnt pathway, including arm, in the MB of adult flies disrupted long term memory in the olfactory behavioral paradigm [35]. Finally, HUWE1 was identified as an interactor of the ubiquitin E3 ligase RNF146, which was demonstrated to promote Wnt signaling [36].

We could ameliorate the increased branching phenotype caused by HUWE1 overexpression upon co-expressing a constitutively active form of the β-catenin homolog arm, indicating that the canonical Wnt/β-catenin pathway is affected by increased HUWE1 levels. However, as the phenotype was not completely rescued, other Wnt signaling pathways might play a role as well. It is of significant interest to note that two nonsense and one frameshift mutation in the CTNNB1 gene that encodes the β-catenin protein have recently been described in patients with very similar clinical features including severe ID with absent or very limited speech, microcephaly and spasticity [37]. Regarding signaling, we can exclude involvement of JNK signaling, as the dsh mutations, which is specifically mutated in the DEP domain necessary for activation of JNK signaling [21], did not affect terminal axon branching. The Wnt-dsh-JNK signaling pathway has been shown to regulate axon extension and retraction in the DCNs, and both dsh*mutants affect this pathway causing a reduction in the number of DCN axons reaching the medulla [20]. Although western blot indicated that HUWE1 reduces the dsh protein levels, we did not see an effect on the DCN axon numbers in the medulla. The effect of HUWE1 on terminal branching without affecting axon outgrowth via JNK signaling could be explained by the existence of different dsh pools, with HUWE1 possibly selectively affecting one pool without targeting the other. Dvl/dsh has been shown to shuttle between the cytoplasm and the nucleus [38-42], and interestingly, disruption of the nuclear localization signal of dsh specifically impairs the canonical β-catenin pathway [41]. Another study indicated that this nuclear Dvl is crucial for the formation of a stable complex between β-catenin and T cell factor (TCF), via which it can affect β-catenin-dependent transcription [39]. If overexpression of HUWE1 specifically affects the nuclear dsh pool, this could account for the absence of non-canonical pathway-dependent phenotypes. Alternatively, the axon branching could be more sensitive to changes in dsh levels compared to the axon extension. The fact that dsh RNAi flies...
still have on average 11.3 axons projecting to the medulla is in agreement with this hypothesis.

To conclude, increased expression of HUWE1 did not cause major structural defects in the brain of Drosophila, but the terminal axon branching of the DCNs was severely disturbed. Our results suggest that this branching phenotype is caused by enhanced breakdown of dsh through increased levels of HUWE1 resulting in reduced activation of the canonical Wnt/β-catenin pathway. It therefore is tempting to speculate that a similar process is acting in ID patients with duplication of HUWE1, linking Wnt/β-catenin signaling to memory formation.

Supporting Information

Figure S1. Automated analysis of the NMJ. Visualization of the NMJ analysis as generated by an in-house developed ImageJ/Fiji-based macro. Anti-Dig1 staining is shown in red. The blue line traces the length and branches of the NMJ, whereas the yellow line on the outside of the NMJ visualizes the NMJ perimeter. The white dots represent the active zones (as determined by anti-nc82 staining).

Table S1. Primers used for PCR and RT-qPCR.

Table S2. Quantification of NMJ parameters in 25 controls and 30 HUWE1 transgenes. NMJ parameters were automatically quantified by an in-house developed ImageJ/Fiji-based macro, as visualized in Figure S1. NMJ area was normalized to muscle area. No parameters reached a p-value < 0.01 (Student's t-test).

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Author Contributions

Conceived and designed the experiments: JV PV AS BAH GF. Performed the experiments: JV ML MZ HB BN EL MS. Analyzed the data: JV HB JMK MB. Contributed reagents/materials/analysis tools: PM AS. Wrote the manuscript: JV AS BAH GF.

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Increased HUWE1 in Flies Causes Branching Defects

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