Diversity and Ecophysiology of New Isolates of Extremely Acidophilic CS\textsubscript{2}-Converting *Acidithiobacillus* Strains


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Biofiltration of industrial carbon disulfide (CS\textsubscript{2})-contaminated waste air streams results in the acidification of biofilters and therefore reduced performance, high water use, and increased costs. To address these issues, we isolated 16 extremely acidophilic CS\textsubscript{2}-converting *Acidithiobacillus* thiooxidans strains that tolerated up to 6% (vol/vol) sulfuric acid. The ecophysiological properties of five selected strains (2Bp, Sts 4-3, S1p, G8, and BBW1) were compared. These five strains had pH optima between 1 (2Bp) and 2 (S1p). Their affinities for CS\textsubscript{2} ranged between 80 (G8) and 130 (2Bp) μM. Strains S1p, G8, and BBW1 had more hydrophobic cell surfaces and produced less extracellular polymeric substance than did strains 2Bp and Sts 4-3. All five strains converted about 80% of the S added as CS\textsubscript{2} to S\textsubscript{0} when CS\textsubscript{2} was supplied in excess. The rate of S\textsubscript{0} consumption varied between 7 (Sts 4-3) and 63 (S1p) nmol O\textsubscript{2} min\textsuperscript{-1} ml culture\textsuperscript{-1}. Low S\textsubscript{0} consumption rates correlated partly with low levels of cell attachment to externally produced S\textsubscript{0} globules. During chemostat growth, the relative amount of CS\textsubscript{2} hydrolase in the cell increased with decreasing growth rates. This resulted in more S\textsubscript{0} accumulation during CS\textsubscript{2} overloads at low growth rates. Intermittent interruptions of the CS\textsubscript{2} supply affected all five strains. Strains S1p, G8, and BBW1 recovered from 24 h of starvation within 4 h, and strains 2Bp and Sts 4-3 recovered within 24 h after CS\textsubscript{2} was resupplied. We recommend the use of mixtures of *Acidithiobacillus* strains in industrial biofilters.

Carbon disulfide (CS\textsubscript{2}) is a toxic, volatile, flammable, and explosive solvent widely used in, e.g., the viscose rayon industry (1). Because of its toxicity and the increasingly stringent rules governing the emission of harmful gases, it is necessary to treat CS\textsubscript{2}-containing industrial waste gases. Biological treatment of CS\textsubscript{2} (and hydrogen sulfide, H\textsubscript{2}S) with sulfur-oxidizing bacteria provides an attractive alternative to conventional treatment systems (e.g., active carbon, incineration, caustic scrubbing) (2, 3). Typical concentrations of CS\textsubscript{2} in contaminated air from viscose industries are around 4 to 20 nmol ml\textsuperscript{-1} (100 to 500 ppm) (2).

The number of microorganisms known to be able to grow chemolithoautotrophically on CS\textsubscript{2} is limited to some *Thiobacillus* species, *Thiotrichia ramosa*, *Paracoccus denitrificans*, and a *Thiomasia* sp. (4–8). All of these CS\textsubscript{2}-utilizing bacteria grow at neutral pH. Thus far, only one CS\textsubscript{2}-utilizing species (*Thiobacillus* sp. strain TJ330, DSM8985) capable of growth under acidic conditions (as low as pH 0.5) has been described (9). The only reported screening of 10 (*Acidithiobacillus*) strains showed that CS\textsubscript{2} conversion is not a general trait of (*acidit*)hiothiobacillus (4). In that screening, only one strain, *Thiobacillus thioparus* TK-m, was found to be capable of CS\textsubscript{2} conversion. We recently discovered that CS\textsubscript{2} conversion is not limited to the domain *Bacteria*; the hyperthermoacidiphilic archaea *Acidianus* sp. strain A1-3 and *Sulfolobus solfataricus* P2 can also grow on CS\textsubscript{2} as a main carbon and energy source (10). However, these archaea are not able to grow at the extremely low pH values that acidithiobacilli can cope with.

CS\textsubscript{2}-converting sulfur oxidizers in operating biofilters are acidophilic bacteria (2, 11–13). They convert CS\textsubscript{2} via the two hydrolysis reaction steps CS\textsubscript{2} + H\textsubscript{2}O → COS + H\textsubscript{2}S and COS + H\textsubscript{2}O → CO\textsubscript{2} + H\textsubscript{2}S and obtain their energy from the oxidation of H\textsubscript{2}S via S\textsuperscript{0} and SO\textsubscript{2}\textsuperscript{−} to SO\textsubscript{4}\textsuperscript{2−} as follows: H\textsubscript{2}S + SO\textsubscript{2}\textsuperscript{−} → SO\textsubscript{4}\textsuperscript{2−} + 2H\textsuperscript{+} (14). Therefore, an inherent result of CS\textsubscript{2} conversion is acidification of the biofilters, which can be limited only by flushing the trickling filters with fresh water. Operating at a pH as low as possible will considerably reduce the volume of fresh water used for neutralization. Water use would be further reduced if the H\textsubscript{2}SO\textsubscript{4} produced could be reused in the viscose-rayon industry. This becomes economically feasible when the H\textsubscript{2}SO\textsubscript{4} concentration in the reactor effluent is at least 10% (wt/vol) (5.6% [vol/vol]). However, the performance of biotrickling filters is compromised by severe acidification and buildup of elemental sulfur (S\textsuperscript{0}) that can clog the filters. Therefore, we set out to isolate new CS\textsubscript{2}-converting bacterial strains able to tolerate extremely low pH values with variable CS\textsubscript{2} loads without loss of CS\textsubscript{2} conversion efficiency and without the production of large amounts of elemental sulfur.

**MATERIALS AND METHODS**

**Media and culture conditions.** Strains were enriched and cultured in basal salt mineral medium (MM) with CS\textsubscript{2} as the sole carbon and energy source as described previously (15). Bacteria were grown at room temperature (RT, 22°C) in 120-ml bottles containing 20 ml MM acidified with sulfuric acid. Alternatively, bacteria were grown on MM plates solidified with 1% (vol/vol) Gelrite (16) and acidified with 0.1% (vol/vol) sulfuric acid. This was the maximum [H\textsubscript{2}SO\textsubscript{4}] at which plates could still be poured without the Gelrite solution immediately solidifying when brought into contact with the H\textsubscript{2}SO\textsubscript{4}-containing MM solution. In the case of Gelrite, H\textsubscript{2}SO\textsubscript{4} increases solidification while the opposite occurs with agar(ose). Plates were incubated in an airtight jar. Unless stated differently, sulfuric acid. Alternatively, bacteria were grown on MM plates solidified with 1% (vol/vol) Gelrite (16) and acidified with 0.1% (vol/vol) sulfuric acid. This was the maximum [H\textsubscript{2}SO\textsubscript{4}] at which plates could still be poured without the Gelrite solution immediately solidifying when brought into contact with the H\textsubscript{2}SO\textsubscript{4}-containing MM solution. In the case of Gelrite, H\textsubscript{2}SO\textsubscript{4} increases solidification while the opposite occurs with agar(ose). Plates were incubated in an airtight jar. Unless stated differently, sulfuric acid.

**Received** 1 July 2013 **Accepted** 25 August 2013

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equivalent to 1.8% [wt/vol] and 0.18 M sulfuric acid). Strains were also grown in minichemostat reactors as described previously (15).

The headspaces of the bottles and jars used were continuously flushed with a CS₂-containing air stream from a purpose-built distribution system (see the supplemental material).

**Enrichment and isolation.** Volumes of 0.5 to 1 ml of environmental or industrial samples were inoculated into 120-ml serum bottles with 20 ml acidified MM. The initial H₂SO₄ concentration was 0.5 to 1% (vol/vol) in the environmental samples and 2% in the samples from the biotrickling filters. CS₂ was supplied as the sole energy source via the distribution system. When a visually dense culture was obtained, the enrichment was transferred to fresh MM. The maximum H₂SO₄ concentration at which growth occurred was determined by the subsequent transfer of enrichments to MM with higher H₂SO₄ concentrations. Pure cultures were obtained from enrichment cultures grown in 4 to 6% H₂SO₄ on Gelrite plates. Single colonies were serially transferred three times to fresh plates and checked microscopically for purity. Fungal contamination (present mainly in the enrichments from industrial samples) was eliminated by adding 150 μg ml⁻¹ chlorothalonil either to the plates or to liquid cultures. The H₂SO₄ tolerance of each isolated strain was confirmed by repeated subculturing at least three times in liquid MM containing 4 to 5% H₂SO₄.

**Screening of known Acidithiobacillus strains for CS₂ conversion capacity.** Five Acidithiobacillus strains were obtained from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ) and grown in batch cultures in the media suggested (DSMZ medium number in parentheses after the strain name) or as otherwise stated, i.e., *A. ferrooxidans* DSM14882 (882), *A. caldus* DSM8584 (150a), *A. thiooxidans* DSM14887 (71), *A. thiooxidans* DSM504 (MM plus 10 g liter⁻¹ sterile S²⁻), and *A. albertensis* DSM14366 (71). After growth for 1 week, the headspace of the cultures was supplemented with CS₂ (20 to 30 nmol ml⁻¹). The CS₂ concentration and the presence of intermediates (H₂S and COS) in the headspace were monitored over time by gas chromatography (17).

**PCR, cloning, and sequencing.** DNA was isolated from each strain by phenol extraction (18). The 16S rRNA gene and the 16S-23S intergenic spacer region (ISR) of 16 isolated CS₂-hydrolyzing bacterial strains were amplified by hot-start PCR with the Go Taq Green buffer system (Fermentas) with 2.5 mM MgCl₂, 0.2 mM deoxynucleoside triphosphates, 1 μM bovine serum albumin, 0.4 μM each primer (see Table S1 in the supplemental material), and 1 μM Taq polymerase (Fermentas). The PCR protocol consisted of 2 min at 95°C, 30 cycles of 1 min at 95°C, 1 min at 54 to 65°C, and 2 min at 72°C; and a final elongation step of 10 min at 72°C. PCR products were ligated into pGEM-T Easy (Promega) and transformed into Escherichia coli strain TOP10 (Invitrogen) according to the manufacturers’ instructions. Plasmids with a correct insert were Sanger sequenced by the sequencing facility at the Department of Human Genetics, University of Michigan, Ann Arbor. A total of 48 clones were isolated by plating on agar containing 1 mmol CS₂ l⁻¹ for 24 to 48 h. The PCR products were sequenced on an ABI3730XL automatic sequencer. The sequences were assembled with SeqMan II software (DNASTAR). The sequences were aligned using the ClustalW method, and a phylogenetic tree was generated using the neighbor-joining method in Mega 4.0 (20) with the following settings: maximum composite likelihood nucleotide substitution model, gaps and missing data eliminated, transitions and transversions included, uniform rates among sites, and homogeneous pattern among lineages. Sequences of the ribosomal operon of *A. thiooxidans* ATCC 19377 and *A. caldus* ATCC 51756 were kindly made available by Jorge Valdés and David Holmes, CBGB, Santiago, Chile (21).

**Cryo-SEM.** Cryo-scanning electron microscopy (cryo-SEM) was used to study the morphology of colonies growing on 1% Gelrite plates acidified with 0.018 M H₂SO₄ by a method similar to that described in reference 22. Blocks containing colonies were cut out of the Gelrite, mounted on electroconductive aqueous colloidal graphite (DAG; Agar Scientific) on a mounting stub, and quickly frozen by submersion in degassed liquid N₂ (−196°C). While under vacuum, the sample was transferred to the Gatan cryotransfer box. Sections through colonies were made with a razor blade at this stage when required. The temperature was subsequently increased to −100°C to sublime off water that had settled on top of the specimen by condensation for a period of 5 min. When all of the surface water had been removed, the temperature was reduced to between −100 and −150°C. The sample was then sputter coated for 45 s with a mixture of 60% gold and 40% palladium and transferred to a JEOL 6330F scanning electron microscope.

**Yield determinations.** For total carbon measurements, 3- to 4-ml reactor samples were centrifuged and the pellets were resuspended in 2 ml 1 mM HCl, pH 3. The washed cells were dried overnight under vacuum at 70°C. The C/N ratio of the dried material (0.3 to 0.4 mg) was determined by elemental analysis with a Thermo Fisher Scientific EA 1110 CHN element analyzer coupled to a Finnigan DELTAplus mass spectrometer. For protein determinations, 2-ml reactor samples were centrifuged at 4°C for 30 min at 18,000 × g. The pellets were resuspended in 0.5 ml 1 M NaOH, boiled for 5 min, and neutralized with 0.5 ml 1 M HCl. Alternatively, 0.5 ml 1 M NaOH was added directly to 200-μl reactor samples, and after boiling, the mixture was neutralized with 0.3 ml 1 M HCl. Protein concentrations were determined with the Bio-Rad protein assay kit according to the manufacturer’s instructions. Culture density, determined by both cell density and the presence of S³⁻, was measured spectrophotometrically as the optical density at 600 nm (OD₆₀₀).

**Determination of pH optima.** pH optima were determined by a floating-filter method. Samples taken from the steady-state chemostats (1% H₂SO₄ measured pH, 0.72) were diluted 10⁶-fold in MM at pH 2, and 1 ml of this dilution was filtered through a sterile, 0.2-μm, 25- or 47-mm diameter Cyclopore polycarbonate filter (Whatman). Filters were then floated on 20 ml MM acidified with sulfuric acid to pH 2.5 to 6. They were placed in airtight jars and incubated at RT for 16 days with a continuous flow of 45 ml min⁻¹ air containing 10 nmol CS₂ ml⁻¹. Growth at different pHs was determined by measuring colony diameters and counting colonies. The growth of strain BBW1 was quantitated visually, as cells had spread over the filter and over the surface of the medium.

**Cell surface hydrophobicity.** Cell surface hydrophobicity was determined by a modified form of the method of Rosenberg et al. (23). The pH of culture samples from the minichemostats was adjusted to 3 or 7 with PUM buffer (23). The suspensions were diluted to an OD₆₀₀ of 0.45 (A₆₀₀) in PUM buffer with the appropriate pH. In a test tube, a 1-ml suspension was mixed with 200 μl n-octane or n-hexane. The mixture was incubated at RT for 10 min, mixed vigorously for 1 min, and left to stand at RT for at least 25 min. OD₆₀₀ was measured (A₆₀₀), and the percent adherence to the solvent was calculated with the equation (1 − A₆₀₀/A₆₀₀) × 100. The use of OD₆₀₀ to measure suspension turbidity yielded similar results.

**Preparation of cell extracts.** Cell extracts from steady-state reactor-grown bacterial cells were prepared as follows. Thirty to 50 ml was removed from the reactors and centrifuged at 4°C for 30 min at 12,000 × g. The cell pellets were washed with 15 ml sterile distilled H₂O and resuspended in 0.5 ml 20 mM KP, pH 7. Approximately 350 μM glass beads (80- to 110-μm size) were added, and the cells were broken by bead beating for 2 × 2 min at 30 Hz (Retsch) with intermittent cooling on ice. The broken cell mixtures were centrifuged for 5 min at 16,000 × g, and the supernatants were stored at −20°C with a final concentration of 10% glycerol.

**Enzyme kinetics based on H₂S measurements.** The Michaelis-Menten constants *Kₘ* and *Vₘₐₓ* were determined for the CS₂ conversion rates of cell extracts of the different strains by measuring the H₂S production rate with an H₂S microsensor (Unisense) in 20 mM HEPES (pH 7) as described previously (15). Experiments were performed at pH 7, as the CS₂ hydrolase is predicted to reside in the cytoplasm of the cell because of the absence of a signal sequence at the N terminus of the enzyme. The *Kₘ* and *Vₘₐₓ* values were calculated from Michaelis-Menten plots by nonlinear regression with the Michaelis-Menten equation *V* = *Vₘₐₓ* × [CS₂]/(*Kₘ₄* + [CS₂]). Experiments were repeated at least three times, and average values of three independent experiments ± the standard errors of the means were calculated. The same method was used to determine *Vₘₐₓ* values (with 600 μM CS₂ as the substrate) for steady-state cells from the
minichemostats (D = 0.02) diluted 200× in reactor medium (MM containing 1% H$_2$SO$_4$). Rates were normalized to the reactor OD$_{600}$.  

**Sulfur (S$_0$) determination.** Sulfur was determined by a modified form of the method of Sorbo (24) to reduce interference from medium components and loss of sulfur during processing. Polycarbonate filters (0.1-μm Whatman Cyclopore track etched membrane 7060-2501 or Millipore Isopore VCPT filters) were rinsed with MilliQ water and placed on a vacuum filter unit. Liquid samples from batch and continuous cultures, as well as bacteria removed from Gelrite plates and resuspended in 0.5 ml sterile demineralized water, were immediately filtered to prevent the bacteria from metabolizing the S$_0$ before processing. Up to 5 ml culture was carefully loaded directly onto the membrane to prevent S$_0$ from sticking to the glass of the vacuum unit and filtered under vacuum. Controls consisted of 3 ml MM containing 1% H$_2$SO$_4$. The filters were rinsed with 1 ml MilliQ water and inserted into a 2-ml Eppendorf tube, and 1.5 ml 0.1 M KCN was added immediately to stop the cells from metabolizing the S$_0$. The samples were incubated at 90°C for 10 min and cooled to RT, and 200 μl 0.75 M FeNO$_3$ in 20% HNO$_3$ was added. Samples were centrifuged for 5 min at 16,000 × g to pellet precipitate cell debris and filters, and the supernatant absorbance was measured immediately at a wavelength of 460 nm. Standard curves were prepared in the same manner, with a solution of S$_0$ in acetone (259 mg in 100 ml) diluted in MilliQ water to a final amount of up to 75 μg S$_0$/per filter.

S$_0$, H$_2$S, and SO$_3^{2-}$ accumulation in bacterial cells. The accumulation of intermediates during CS$_2$ respiration was measured simultaneously but in separate reaction chambers as follows. Samples from the minichemostats were diluted 10-fold with O$_3$-saturated MM containing 1% H$_2$SO$_4$. The diluted cultures were used to fill three glass cuvettes containing stir bars, after which the cuvettes were closed and placed in a 22°C water bath for 35 min with vigorous stirring. CS$_2$ (17 μM) was added to each cuvette, and simultaneous measurements were then made of (i) O$_2$ respiration with an O$_2$ sensor (Unisense), (ii) H$_2$S production with an H$_2$S sensor (Unisense), and (iii) S$_0$ production by following the change in OD$_{480}$ with an Agilent 8453 spectrophotometer. In addition and simultaneously, a 100-ml glass syringe containing a stir bar was filled with diluted culture and incubated in a 22°C water bath for 35 min with continuous stirring. CS$_2$ was also added to the syringe, and at intervals during the respiration and H$_2$S production curves, samples were pushed out of the syringe into an Eppendorf tube and frozen immediately for subsequent S$_0$ determination.

In order to study H$_2$S and S$_0$ formation kinetics during CS$_2$ respiration in more detail, these parameters were also simultaneously measured in one cylindrical cuvette with three entry ports, which was positioned in a single-beam spectrophotometer. Two ports were used for O$_2$ and H$_2$S sensors, and the third served for the addition of CS$_2$. The percentage of S$_0$ accumulated was calculated as follows. The total O$_2$ consumption is represented by the formula H$_2$S + 2O$_2$ → H$_2$SO$_4$. In a respiration curve, the total O$_2$ consumed (A) will depend on the total amount of H$_2$S (produced from CS$_2$) and so, A/2 represents the amount of sulfide oxidized to sulfate. Sulfur S$_0$ is an intermediate and is oxidized according to the formula S$_0$ + 1.5O$_2$ + H$_2$O → H$_2$SO$_4$. At the sulfur peak (A$_{100}$), only S$_0$ is present and the amount of oxygen consumed (a) then until the end represents sulfur oxidation, and so, a/1.5 represents the amount of S$_0$ present at the peak. This gives a S$_0$/total sulfide ratio of [a/1.5] × [A/2] = 1.33 × a/A, where a/A is the fraction of the oxygen consumption after the sulfur peak compared to the total consumption, determined from the respiration curve.

**Sulfite (SO$_3^{2-}$) determination.** Sulfite (SO$_3^{2-}$) was measured by the method described by Truex and Schlegel (25) but with reduction of the amount of H$_2$SO$_4$ in the assay to take into account the amount of H$_2$SO$_4$ that is present in the samples (in these experiments, 1% [vol/vol]). Dilutions of an anaerobic 1 M stock of Na$_2$SO$_3$ were used as standards.

**Starvation experiments.** Starvation experiments were performed in steady-state chemostats by stopping the influent and effluent pumps and removing the CS$_2$ supply. One-milliliter samples were removed at regular intervals from the reactors during starvation and recovery, diluted 10× in MM containing 1% (vol/vol) H$_2$SO$_4$ and incubated in a 7.63-ml double-port cuvette at 22.0 to 22.2°C. Ten minutes after the sample was removed from the chemostat, a 10 μM CS$_2$ pulse from a 6 ml stock bottle (see above) was injected into the cuvette. H$_2$S production and removal, as well as respiration, were measured simultaneously with an H$_2$S microsensor (Unisense) and an oxygen sensor (Strathkelvin Instruments), respectively.

**Nucleotide sequence accession numbers.** The 16S ISR sequences determined in this research have been deposited in the GenBank database under accession numbers KC902816 to KC902829 and KC902831.

**RESULTS**

**Enrichment and isolation of CS$_2$-utilizing bacteria.** To obtain extremely acidophilic CS$_2$-converting microorganisms, samples from naturally acidic, sulfur-rich environments and from industrial biotrickling filters were incubated in acidified MM and with CS$_2$ as the sole energy source. Significant CS$_2$ conversion was observed within a few days, and dense cultures were obtained within 2 to 4 weeks of incubation with 1 to 4% H$_2$SO$_4$. Ultimately, some enrichment cultures showed growth at 6% H$_2$SO$_4$ (Table 1). This concentration is equal to a theoretical pH of ~0.05. Enrichments containing 4 to 6% H$_2$SO$_4$ (Table 1) were used to isolate 16 pure cultures on Gelrite plates (0.1% H$_2$SO$_4$). To reconfirm their acid tolerance, all isolates were successfully transferred to 4% H$_2$SO$_4$ medium.

Descriptions of growth characteristics on plate cultures and in liquid cultures are given in the supplemental material and shown in Fig. S1 and S2 in the supplemental material. The 16 isolated strains showed distinctly different colony morphologies, ranging from large, dry, white colonies to smooth, shiny, compact colonies. Stationary-phase liquid cultures showed white or yellow aggregates or little aggregation. From the cryo-SEM analysis of surfaces and cross sections of colonies, we conclude that on solid medium, the compactly growing strains produce more extracellular polymeric substance (EPS) than the spreading strains do.

**Phylogenetic analysis.** All of the new CS$_2$-converting isolates were identified as Acidithiobacillus thiooxidans strains by conventional 16S rRNA gene analysis (data not shown). Improved discrimination was achieved by analysis of the 16S-23S ISR sequences, containing two tRNA genes and three intergenic transcribed spacers (ITS). The total length of the ISR sequences of the CS$_2$-converting isolates varied between 456 and 460 bp, with one exception; isolate BDW2 from the Oy Visko reactor had an ISR of 439 bp (see Table S2 in the supplemental material). Within the ISR, most of the variation in nucleotide composition was observed in the third of the three ITS. Strains BAW3 and BBW1, which originated from two parallel reactors operating at the Loudon factory, were distinctly different with respect to their nucleotide composition, differing by 57 bp and harboring three insertions and three deletions compared with reference strain G8. In a neighbor-joining phylogenetic tree constructed from the 16S ISR sequences of the CS$_2$-converting strains and other A. thiooxidans strains, strains BAW3 and BBW1 form a distinct cluster within the A. thiooxidans strains (Fig. 1). Three more clusters could be distinguished, i.e., cluster 2, which did not contain any of the CS$_2$-converting strains; cluster 3, comprising most of the CS$_2$-converting strains and also A. albertensis, described as a distinct species but phylogenetically indistinguishable from A. thiooxidans (26); and cluster 4, containing strain S1p from the Solfatara
(Rome, Italy), strain BC6-1, and strain BDW2 from the Oy Visko plant.

**Screening of Acidithiobacillus reference strains for CS2 conversion.** To check whether CS2 conversion is a general trait of acidithiobacilli under our growth conditions, we tested four publicly available Acidithiobacillus species. Of these, the three mesophilic Acidithiobacillus species (A. albertensis DSM14366, A. ferrooxidans DSM14882, and A. thiooxidans DSM504 and DSM14887) were not able to convert CS2 during the 2 to 5 days the cultures were monitored. Since a hierarchical utilization of energy sources may still be possible, CS2 utilization cannot be completely ruled out. However, the moderate thermophile Acidithiobacillus albertensis DSM4684 was found to be capable of CS2 conversion as soon as CS2 was added to a growing culture.

**Maximum acid tolerance.** The maximum acid tolerance of all 16 isolates in batch cultures was determined by serially transferring the strains to medium containing gradually higher concentrations of H2SO4. The maximum H2SO4 concentration in which growth could be observed within 2 months was taken as the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism. Table 1 shows that all of the isolated strains were able to grow at 4% H2SO4, which corresponds to the growth limit of each organism.
more acid (pH 0.5) and more gradually decreased when cells were exposed to higher-than-optimum pHs. Strain BBW1 was the most sensitive to high-pH stress; no growth was observed in medium with a pH higher than 3.5.

**Enzyme kinetic analysis.** To assess the CS$_2$ removal efficiencies of the five new CS$_2$-converting *A. thiooxidans* isolates, the CS$_2$ affinity constants ($K_m$) and maximum CS$_2$ conversion rate ($V_{max}$) of crude protein extracts from continuous cultures were determined by adding a pulse of CS$_2$ to diluted protein extracts and measuring the rate of H$_2$S production. Table 2 shows that the $K_m$ values of the five strains are of the same order of magnitude, ranging between 81 and 130 $\mu$M for strains G8 and 2Bp, respectively. Comparison of the $V_{max}$ values revealed crude extracts from strain S1p to have a consistently higher $V_{max}$ under these conditions than crude extracts from the other four strains tested, resulting in a 2-fold higher $k_{cat}/K_m$ value for S1p. In accordance, the $V_{max}$ for CS$_2$ of steady-state, minichemostat-grown, intact S1p cells was consistently slightly higher than the $V_{max}$ of the other strains (Table 2).

**Sulfur production and consumption.** (i) Sulfur production. Production of S$^0$ by CS$_2$-converting acidithiobacilli as an intermediate in the oxidation of H$_2$S to SO$_4^{2-}$ can block biotrickling filters and subsequently decrease performance. Therefore, we compared the H$_2$S and S$^0$ production and consumption of the five selected *A. thiooxidans* strains upon CS$_2$ pulses. When concentrations as low as 1.8 $\mu$M CS$_2$ (equivalent to 36 ppm CS$_2$ in the gas phase) were added to a cuvette containing diluted samples from the minichemostats, there was an almost immediate increase in S$^0$, just above the detection level. Peak concentrations of H$_2$S ranged from 0.5 to 1.0 $\mu$M under these conditions, and so, S$^0$ formation already started at H$_2$S concentrations below this value. In subsequent experiments, H$_2$S and S$^0$ production was measured simultaneously in one cuvette. Addition of 17$\mu$M CS$_2$ resulted in an immediate and large accumulation of H$_2$S for all strains. Also, S$^0$ formation started almost without delay and continued until virtually all of the H$_2$S had been consumed (Fig. 2). The respiration curves show an initial period of fast respiration when H$_2$S is still present, followed by a sudden decrease in the respiration rate when the cells start respiring solely on S$^0$. This decrease in the respiration rate indicates that the processing of the S intermediates in the cell, and not respiration, is the rate-limiting step. This was observed in all of the strains, but it was most prominent in strain Sts 4-3 (Fig. 2). The chemical reaction of H$_2$S with O$_2$ was less than 2 $\mu$M H$_2$S h$^{-1}$ and therefore did not significantly contribute to the observed oxygen consumption rates.

During the period of S$^0$ respiration, we often observed a temporary reduction in the respiration rate. Although it is possible that an intermediate temporarily accumulated that inhibited res-

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**FIG 1** Phylogenetic tree based on the 16S-23S ISR of CS$_2$-converting *A. thiooxidans* strains isolated from environmental samples (1Bp, 2Ap, 2Bp, Sts 4-3, and S1p) or various industrial CS$_2$ biofilter effluents (G8, BAD2, BED2, BBF2, BBW1, BAW3, BEF1, BDW2, BC6-1, and BEF3). The optimal neighbor-joining tree with a branch length sum of 0.98269283 is shown. The tree is drawn to scale, with branch lengths in the same units as the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed by the maximum composite likelihood method and are expressed as the number of base substitutions per site. All positions containing gaps and missing data were eliminated. There were a total of 330 positions in the final data set. The percentage above 50% of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) is shown next to the branches. Four clusters of *A. thiooxidans* strains were identified, based on branch points that were reproduced in more than 50% of the bootstrap replicates. The type strains are *A. thiooxidans* ATCC 19377, *A. albertensis* DSM14366, and *A. caldus* DSM8584. In bold and underlined are strains that form white, large, dry, crusty colonies on Gelrite plates. In bold are strains that form small, shiny, smooth colonies on Gelrite plates.
pitation on S\textsuperscript{0}, at this point, the S\textsuperscript{0} consumption rate did not decrease. Also, although detectable levels of the potentially inhibiting intermediate SO\textsubscript{4}\textsuperscript{2-} were present throughout the experiment, there was no increase at the reduced respiration “bump.” Therefore, the apparent temporary reduction in respiration is more likely due to the sensor being affected by an intermediate at that point in the experiment.

At the point where S\textsuperscript{0} peaks, the H\textsubscript{2}S and CS\textsubscript{2} had been completely consumed and only 30 to 40% of the total oxygen was consumed. Therefore, the remaining O\textsubscript{2} consumption resulted entirely from S\textsuperscript{0} oxidation, if the accumulation of any other intermediates is excluded. With this O\textsubscript{2} consumption, and taking into account that this is an underestimate of the actual O\textsubscript{2} consumption because of a small baseline drift upward of the O\textsubscript{2} sensor during the experiment, we calculated that at least 80% of the S added as CS\textsubscript{2} accumulated as S\textsuperscript{0}. This was similar for all of the strains tested (Table 2), and the percentage also remained identical when CS\textsubscript{2} concentrations as low as 1.8 μM were added. Below this value, H\textsubscript{2}S formation could not be observed any more, making such calculations impossible.

The rates of S\textsuperscript{0} formation and consumption were calculated from the slopes before and after the apex of the OD\textsubscript{480} trace (Fig. 2). Consistently, S\textsuperscript{0} production (corrected for consumption) was between four times (strains 2Bp, S1p, G8, and BBW1) and nine times (strain Sts 4-3) faster than S\textsuperscript{0} consumption. This matches with a buildup of S\textsuperscript{0} of at least 80% of the total S added, as calculated above from the oxygen respiration. In separate experiments, chemical analysis for S\textsuperscript{0} at the moment OD\textsubscript{480} reached its maximum value consistently showed S\textsuperscript{0} recovery between 60 and 67% (Table 2). Therefore, chemical analysis appeared to underestimate the amount of S\textsuperscript{0} produced during CS\textsubscript{2} respiration. However, both methods of determining S\textsuperscript{0} accumulation upon the application of CS\textsubscript{2} pulses indicated that all of the strains tested produced equal amounts of extracellular S\textsuperscript{0} under conditions of excess CS\textsubscript{2}.

(ii) Sulfur consumption. Although all of the strains tested behaved similarly in terms of S\textsuperscript{0} production, there were some obvious differences in subsequent S\textsuperscript{0} respiration (Fig. 2); strain Sts 4-3 respired on S\textsuperscript{0} five to eight times more slowly than the other four strains. The H\textsubscript{2}S combined with S\textsuperscript{0} respiration rates (the first part of the respiration curve) showed a maximum 2-fold difference between the strains (Fig. 2 and Table 2). Although strain Sts 4-3 respired more slowly on S\textsuperscript{0}, the respiration rate did not decline until all of the S\textsuperscript{0} had been depleted.

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**TABLE 2** Comparison of CS\textsubscript{2}-converting *A. thiooxidans* strains 2Bp, Sts 4-3, S1p, G8, and BBW1a

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>2Bp</th>
<th>Sts 4-3</th>
<th>S1p</th>
<th>G8</th>
<th>BBW1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Colony morphology</td>
<td>Smooth, domed</td>
<td>Small, smooth, domed</td>
<td>White, dry, compact</td>
<td>White, dry, spreading</td>
<td>White, dry, spreading</td>
</tr>
<tr>
<td>A. thiooxidans 16S ISR cluster (Fig. 1)</td>
<td>3</td>
<td>3</td>
<td>4</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>Maximum acid tolerance (% [vol/vol] sulfuric acid)</td>
<td>5</td>
<td>5</td>
<td>5</td>
<td>4</td>
<td>6</td>
</tr>
<tr>
<td>pH optimum</td>
<td>1</td>
<td>1.5</td>
<td>2</td>
<td>1.5</td>
<td>1–1.5</td>
</tr>
<tr>
<td>pH tolerance</td>
<td>0.5–6</td>
<td>0.5–5</td>
<td>1–6</td>
<td>0.5–6</td>
<td>0.5–3.5</td>
</tr>
<tr>
<td>Cell surface hydrophobicity</td>
<td>Low</td>
<td>Low-intermediate</td>
<td>High</td>
<td>High</td>
<td>High</td>
</tr>
<tr>
<td>Reactor dilution rate at point of S\textsuperscript{0} formation (approaching μm\textsubscript{max})</td>
<td>0.09</td>
<td>0.08</td>
<td>0.1</td>
<td>0.1</td>
<td>0.09</td>
</tr>
<tr>
<td>Mean cell extract K\textsubscript{m} (μM CS\textsubscript{2}) ± SEM</td>
<td>130 ± 22</td>
<td>97 ± 10</td>
<td>100 ± 5</td>
<td>81 ± 7</td>
<td>116 ± 4</td>
</tr>
<tr>
<td>Mean cell extract V\textsubscript{max} (μmol H\textsubscript{2}S min\textsuperscript{-1} mg protein\textsuperscript{-1}) ± SEM</td>
<td>38 ± 3</td>
<td>17 ± 0</td>
<td>48 ± 7</td>
<td>23 ± 1</td>
<td>28 ± 5</td>
</tr>
<tr>
<td>Mean whole-cell V\textsubscript{max} (μmol H\textsubscript{2}S min\textsuperscript{-1} ml culture\textsuperscript{-1} for OD\textsubscript{600} of 1) ± SEM</td>
<td>11 ± 0</td>
<td>10 ± 0</td>
<td>12 ± 0</td>
<td>8 ± 0</td>
<td>11 ± 0</td>
</tr>
<tr>
<td>Mean calculated S\textsuperscript{0} accumulated (% of total S pulse added) ± SEM</td>
<td>76 ± 4</td>
<td>83 ± 1</td>
<td>82 ± 1</td>
<td>83 ± 1</td>
<td>ND\textsuperscript{b}</td>
</tr>
<tr>
<td>Mean measured S\textsuperscript{0} accumulated (% of total S pulse added) ± SEM</td>
<td>67 ± 2</td>
<td>63 ± 2</td>
<td>65 ± 1</td>
<td>61 ± 1</td>
<td>ND</td>
</tr>
<tr>
<td>Mean respiration rate on H\textsubscript{2}S and S\textsuperscript{0} combined after a 10 μM CS\textsubscript{2} pulse (nmol O\textsubscript{2} min\textsuperscript{-1} ml culture\textsuperscript{-1} for OD\textsubscript{600} of 1) ± SEM</td>
<td>45 ± 9</td>
<td>43</td>
<td>87 ± 10</td>
<td>101 ± 1</td>
<td>66 ± 7</td>
</tr>
<tr>
<td>Mean respiration rate on S\textsuperscript{0} only after 10 μM CS\textsubscript{2} pulse (nmol O\textsubscript{2} min\textsuperscript{-1} ml culture\textsuperscript{-1} for OD\textsubscript{600} of 1) ± SEM</td>
<td>41 ± 3</td>
<td>7</td>
<td>63 ± 3</td>
<td>44 ± 1</td>
<td>49 ± 1</td>
</tr>
<tr>
<td>Mean % of S\textsuperscript{0} globules attached to cells after 10 17 μM CS\textsubscript{2} pulses ± SEM</td>
<td>87 ± 7</td>
<td>17 ± 4</td>
<td>9 ± 2</td>
<td>52 ± 2</td>
<td>22 ± 2</td>
</tr>
<tr>
<td>Mean % of cells attached to S\textsuperscript{0} globules after 10 17 μM CS\textsubscript{2} pulses ± SEM</td>
<td>35 ± 10</td>
<td>13 ± 3</td>
<td>7 ± 2</td>
<td>30 ± 6</td>
<td>13 ± 0</td>
</tr>
<tr>
<td>Resistance to CS\textsubscript{2} stress</td>
<td>Low</td>
<td>Medium</td>
<td>Medium</td>
<td>Medium</td>
<td>Medium</td>
</tr>
<tr>
<td>Speed of recovery after CS\textsubscript{2} starvation</td>
<td>Slow</td>
<td>Slow</td>
<td>Medium</td>
<td>Medium</td>
<td>Medium</td>
</tr>
</tbody>
</table>

\textsuperscript{a} Strains were grown in continuous culture (D = 0.02) with CS\textsubscript{2} as the sole energy source. Samples were taken from these reactors for the comparison experiments. The K\textsubscript{m} and V\textsubscript{max} of each strain were calculated from Michaelis-Menten plots by nonlinear regression with the Michaelis-Menten equation \(V = V_{\text{max}} \times S/(K_{\text{m}} + S)\). Mean values from at least three independent experiments are shown.

\textsuperscript{b} ND, not determined.
This was in contrast to the other strains, especially strain S1p, which showed a continuous decline in the rate of respiration and concurrent S\textsuperscript{0} depletion.

We hypothesized that different strains produce differently bioavailable S\textsuperscript{0}; strains 2Bp, G8, and BBW1 produce S\textsuperscript{0} that can be efficiently metabolized again, whereas strains Sts 4-3 and S1p produce S\textsuperscript{0} that is difficult to remove. To test this hypothesis, we examined sulfur accumulation by different strains upon the application of repeated CS\textsubscript{2} pulses to undiluted chemostat samples by counting S\textsuperscript{0} globules under a light microscope (Table 2). We counted 160 to 240 cells of each strain, and a representative picture of strains 2Bp, Sts 4-3, S1p, and G8 is shown in Fig. S4 in the supplemental material. All five of the strains tested showed the accumulation of S\textsuperscript{0} globules of similar sizes, either attached to the bacteria or loose in the medium (see Fig. S4). There were large differences in the observed cell attachment to S\textsuperscript{0} globules. Although a correlation between the attachment of cells to S\textsuperscript{0} globules and cell surface hydrophobicity might be expected, we did not observe this despite the fact that all of the cells had identical physiological backgrounds. Strain S1p clearly had the fewest S\textsuperscript{0} globules that were attached to cells (9% ± 2%), while in cultures of strain 2Bp, most of the S\textsuperscript{0} globules were attached to cells (87% ± 7%, Table 2). This supports our hypothesis that S1p produces S\textsuperscript{0} that appears less bioavailable; attachment of S\textsuperscript{0} globules to bacteria is required for rapid S\textsuperscript{0} consumption upon H\textsubscript{2}S depletion. Strain Sts 4-3, which consumes S\textsuperscript{0} very slowly, also had a low percentage (13%) of cells attached to S\textsuperscript{0} globules, and 17% of the S\textsuperscript{0} globules were attached to cells (Table 2). This percentage was not as low as that of strain S1p, and it therefore cannot entirely explain the very low observed S\textsuperscript{0} consumption rate of this strain. However, strain Sts 4-3 produced more H\textsubscript{2}S than the other strains when pulsed with CS\textsubscript{2} (25 versus 20 \textmu M, Fig. 2), which could have had an inhibitory effect on overall respiration.

**CS\textsubscript{2} stress and starvation.** The effect of fluctuations in CS\textsubscript{2} concentrations in biofilters can be simulated by applying CS\textsubscript{2} pulses to undiluted culture samples from CS\textsubscript{2}-limited steady-state chemostats in MM with 1% H\textsubscript{2}SO\textsubscript{4}. Therefore, chemostat samples of the newly isolated *A. thiooxidans* strains were incubated and subjected to repeated 17 \textmu M CS\textsubscript{2} pulses. Each new pulse was given when the H\textsubscript{2}S produced from the previous pulse had been
consumed by the cells (after about 1.5 to 3 min). The H$_2$S consumption rates after the first CS$_2$ pulse varied between 1.9 (strain of starvation. After 4 h of CS$_2$ starvation, there was only modest to indicated that the strains cope reasonably well with short periods consumption rates and respiratory activity were measured. Results strain 2Bp, where H$_2$S/S$_0$ respiration activity was reduced by 80% and the subsequent 24-h recovery period when the CS$_2$ supply was reconnected again, samples were taken from the chemostats and the subsequent 24-h recovery period when the CS$_2$ supply was shut down. During this period of starvation and the subsequent 24-h recovery period when the CS$_2$ supply was reconnecting again, samples were taken from the chemostats and subjected to a CS$_2$ pulse. Subsequent H$_2$S production and consumption rates and respiratory activity were measured. Results indicated that the strains cope reasonably well with short periods of starvation. After 4 h of CS$_2$ starvation, there was only modest to no reduction in respiration activity when the cells were pulsed with 10 $\mu$M CS$_2$. Of the four strains tested at this time point, G8 appeared to be the most strongly affected (1.0 $\mu$mol H$_2$S liter$^{-1}$ min$^{-1}$ unit of OD$_{600}^{-1}$, 41% of the initial rate) and strain BBW1 appeared to be the least strongly affected (1.5 $\mu$mol H$_2$S liter$^{-1}$ min$^{-1}$ unit of OD$_{600}^{-1}$, 76% of the initial rate, Fig. 3). This indicates that the cells were becoming increasingly stressed when subjected to CS$_2$ loads above the concentration the cells were adapted to in the chemostat. This is probably due to the toxic effect of the repeated buildup of H$_2$S in the cells.

To test the effect of intermittent interruption of the factory CS$_2$, the five strains growing in chemostats as described above were subjected to CS$_2$ starvation by temporarily shutting down the medium and CS$_2$ supply for 24 h. During this period of starvation and the subsequent 24-h recovery period when the CS$_2$ supply was reconnected again, samples were taken from the chemostats and subjected to a CS$_2$ pulse. Subsequent H$_2$S production and consumption rates and respiratory activity were measured. Results indicated that the strains cope reasonably well with short periods of starvation. After 4 h of CS$_2$ starvation, there was only modest to no reduction in respiration activity when the cells were pulsed with 10 $\mu$M CS$_2$. Of the four strains tested at this time point, G8 appeared to be the most strongly affected, with a 40% reduction of its initial respiration activity, when both H$_2$S and S$_0$ were present and a 22% reduction in S$_0$ respiration activity (Fig. 4). After 24 h of CS$_2$ starvation, all of the strains showed reduced respiratory activity. Although not tested extensively, the reduction was largest for strain 2Bp, where H$_2$S/S$_0$ respiration activity was reduced by 80% and S$_0$ respiration was reduced by 85% below the steady-state culture density of all five strains was higher than during steady state. This indicates that the cultures had started growing and/or had S$_0$ present.

DISCUSSION

Our approach to the enrichment of extremely acidophilic CS$_2$-converting microorganisms from volcanic regions and CS$_2$-converting trickling filters proved successful. Conversion of CS$_2$ was observed within days after inoculation. Furthermore, the enrichments and isolated Acidithiobacillus strains could grow at very high H$_2$SO$_4$ concentrations. The only acidophilic CS$_2$-converting Acidithiobacillus strain described to date, TJ330, was isolated from an acidic (pH 6.1 to 1.4) CS$_2$- and H$_2$S-treated peat biofilter (9). The neutrophilic CS$_2$-converting species Paracoccus denitrificans was isolated from oak leaves and soil beneath the leaf canopy (7, 27). Our attempt to enrich acidophilic microorganisms from soil samples taken under an oak tree and from compost was unsuccessful. The Acidithiobacillus strains from culture collections screened for CS$_2$ conversion (at micromolar concentrations), only A. caldus DSM 8584 was positive. This trait was not previously reported for this species. Our screening confirmed the observation of Smith and Kelly (28), although they used a potentially toxic liquid CS$_2$ concentration (2 mM).

The new Acidithiobacillus isolates even showed slow growth in 6% H$_2$SO$_4$. The constant supply of CS$_2$ to the cultures was the key
factor in the maintenance of cultures at these very high H\textsubscript{2}SO\textsubscript{4} concentrations. Culture activity decreased dramatically upon storage without a substrate at or above 4\% H\textsubscript{2}SO\textsubscript{4}. Acidophiles require a continuous and high supply of maintenance energy to be able to actively pump out protons that leak from the acidic environment into the nearly neutral pH cell cytoplasm (29). Operation of biofilters under highly acidic conditions therefore puts additional stress on the microorganisms during periods of fluctuating CS\textsubscript{2} concentration and factory shutdown.

Although growth was very slow at 6\% H\textsubscript{2}SO\textsubscript{4}, dense growth was already observed at 4\% for all new Acidithiobacillus strains. Similar H\textsubscript{2}SO\textsubscript{4} tolerance was reported for Acidithiobacillus sp. strain AZ11, which is able to respire on elemental sulfur in the presence of 4.2\% H\textsubscript{2}SO\textsubscript{4} (30). The pH optima determined for 5 of the 16 new CS\textsubscript{2}-converting strains were between 1 and 2. This is slightly lower than the optimal-pH range of 1.8 to 2.5 described for other Acidithiobacillus strains (29, 31).

Growth at around pH 0, as observed for 15 of our A. \textit{thiooxidans} isolates, is among the lowest pHs reported in the literature for any organism. \textit{Ferroplasma acidarmanus} (32), \textit{Picrophilus oshimae}, and \textit{Picrophilus torridus} could grow at pH 0 (optimum pH, 0.7) (33). \textit{Picrophilus torridus} adapted to growth at pH 0.1 even showed significant growth at 1.2 M H\textsubscript{2}SO\textsubscript{4} (pH −0.06) (34). The eukaryotic red alga \textit{Cyanidium caldarium} was cultured at 0.5 M H\textsubscript{2}SO\textsubscript{4} (35), and the green alga \textit{Dunaliella acidophila} is able to survive pH 0.2 (36). Also, some fungal species were reported to grow at pH 0, i.e., \textit{Acontium cylatium} (37), \textit{Cephalosporium sp.}, and \textit{Trichosporon cerebratula} (38). We also observed fungal growth at 6\% H\textsubscript{2}SO\textsubscript{4} (pH −0.05) in our enrichment cultures but did not further investigate the species present in these cultures. With growth at pH as low as −0.05 (6\% H\textsubscript{2}SO\textsubscript{4}), the new Acidithiobacillus isolates obtained in this research exceed the previously reported pH limit of 0.5 for microbial CS\textsubscript{2} conversion for Acidithiobacillus TJ330 (9).

The newly isolated CS\textsubscript{2}-converting A. \textit{thiooxidans} strains differed in colony morphology. The genus Acidithiobacillus comprises a physiologically and genetically heterogeneous group of microorganisms (39, 40), despite the often low sequence diversity in the 16S rRNA gene. For that reason, the 16S-23S ISR is used to discriminate at the intra species level (41, 42).

Two main colony types on Gelrite plates were distinguished, compact, creamy, shiny colonies and dry, white, spreading colonies (40). Reversible variation in colony morphology of several \textit{A. ferrooxidans} strains has been described, resulting in the same large, white spreading colonies, as opposed to compact colonies, as observed here for some of the newly isolated A. \textit{thiooxidans} strains (43, 44). A. \textit{thiooxidans} strains may be motile via a polar flagellum (40). A. \textit{ferrooxidans} ATCC 19859 spreading variants displayed increased motility and chemotaxis toward thioulfate, which may be a selective advantage over biofilm growth during periods of low substrate concentrations. These variants arose through the rearrangement of insertion sequences in the genome, potentially acting as a genetic switch (44). The A. \textit{ferrooxidans} group of strains has recently been reclassified into four separate species (45). Of these, only \textit{A. ferrivorans} and some \textit{A. ferridurans} strains were shown to be motile (45). The type strain \textit{A. ferrooxidans} ATCC 23270 lacks flagellum and chemotaxis genes (46). The draft genome of A. \textit{thiooxidans} ATCC 19377 (47) and the draft genomes of both A. \textit{thiooxidans} strains S1p and G8 (Daan Speth, personal communication) do contain the operons for flagellum biosynthesis and chemotaxis. However, we did not observe swimming motility in liquid cultures of our strains when we examined them microscopically. Therefore, the mechanism and role of the spreading colony phenotype in these strains are not clear.

SEM studies of frozen colonies indicated that the difference in colony appearance may be caused by the absence of a clear EPS layer on the dry white colonies, as has been observed for colony morphology mutants of \textit{Mycobacterium smegmatis} (22). In support, A. \textit{thiooxidans} strains S1p, G8, and BBW1, which produced white dry colonies, had a considerably higher cell surface hydrophobicity than strains 2Bp and Sts 4-3, which produced compact shiny colonies, suggesting the presence of relatively more hydrophilic compounds on the cell surface of the latter strains. Acidithiobacillus strains produce EPS containing both neutral sugars and fatty acids (48, 49), the proportion varying depending on the substrate the cells are grown on (50, 51). The EPS of A. \textit{thiooxidans} grown on S\textsubscript{0} consists of 40\% sugars and 60\% fatty acids (mainly eicosanoic acid) (50). The fatty acids are released from the outer membrane by blebbing (52) and cause the “wetting” of sulfur particles described in 1961 by Jones and Starkey (53), making S\textsubscript{0} available as an energy source. In addition, EPS is essential for successful attachment and bioleaching of \textit{A. ferrooxidans} to pyrite (50). Differences in EPS production and potential motility observed in our strains may have implications for the degree of colonization and clogging of biofilters.

Efficient biofiltration of CS\textsubscript{2}-contaminated air streams requires microorganisms with a higher affinity for CS\textsubscript{2} than the concentration present in the air stream, to ensure its removal to concentrations complying with increasingly stringent regulations, as well as resistance to changes in operating conditions, low biomass production, low S\textsubscript{0} accumulation, and rapid S\textsubscript{0} removal to prevent clogging of the biofilters. The affinities of the five strains tested for CS\textsubscript{2} were similar (around 100 \textmu M) and correspond to that reported for the purified CS\textsubscript{2} hydrolase of \textit{Acidithiobacillus} A1-3 (10). Comparison of the \textit{V}_{\text{max}} values revealed strain S1p to have a consistently higher \textit{V}_{\text{max}} under these conditions than the other four strains tested, possibly because it has a relatively larger amount of CS\textsubscript{2} hydrolase present in the cells than the other strains (15). The \textit{V}_{\text{max}} is only reached at substrate concentrations of 200 \textmu M (nmol \textmu M\textsuperscript{-1}) or higher. Bioreactors treating CS\textsubscript{2}-contaminated air from the viscose industry are operational at much lower concentrations of around 4 to 20 nmol \textmu M\textsuperscript{-1} (100 to 500 ppm) (2), indicating that although strain S1p converts CS\textsubscript{2} with a higher \textit{V}_{\text{max}} than the other strains, it would not usually reach its full potential reaction rate in a bioreactor. Indeed, a higher \textit{V}_{\text{max}} may even be deleterious, as it might cause H\textsubscript{2}S to accumulate to toxic levels in the cell more rapidly upon CS\textsubscript{2} peaks.

Comparison of the resistance to changes in operating conditions in the form of an interruption in the CS\textsubscript{2} supply revealed that all five strains were affected by 24 h of CS\textsubscript{2} starvation in terms of the ability the resipe after a CS\textsubscript{2} pulse during starvation, but they all recovered within 24 h after reconnection of the CS\textsubscript{2} supply. However, during recovery, all five strains will also produce S\textsubscript{0}, as we found in all of the strains tested that CS\textsubscript{2} pulses during starvation resulted in the transient accumulation of 80\% of the total S added as S\textsubscript{0}, independently of the amount of CS\textsubscript{2} added. This implies that it will be difficult to avoid S\textsubscript{0} accumulation in biofiltration systems with uneven CS\textsubscript{2} loading. However, differences in S\textsubscript{0} removal were observed, with strain Sts 4-3 having a much lower but constant S\textsubscript{0} removal rate than the other strains and strain S1p.
having a long period in which the $S^0$ respiration rate slowly declined. Whatever the cause, the strains that remove $S^0$ more slowly will cause more clogging problems in bioreactors because of $S^0$ accumulation.

In summary, we have successfully isolated extremely acidophilic, $CS_2$-converting $A. thiioxidans$ strains from both environmental and industrial ecosystems that grow optimally around pH 1 to 2 and can grow at sulfuric acid concentrations of up to 6% (vol/vol). Currently bioreactors are operated at pH 0.5 to 1 (2). Use of the new strains would reduce water use and improve the prospect for reuse of the produced sulfuric acid in the rayon/viscose industry. The isolated strains displayed different growth and colony morphology characteristics, which may be due to differences in motility and/or the presence or absence of an EPS layer surrounding the cells. To circumvent the bottlenecks in the biofiltration of $CS_2$, cocultures of extremely acidophilic $A. thiioxidans$ strains to combine the best acid tolerance, affinity for CS2, $S^0$-removing potential, and stability during periods of fluctuating $CS_2$ loads are most promising for inoculation of industrial biofilters. This application is expected to result in reduced sulfur accumulation, increased $CS_2$ removal rates, reduced water consumption, a more stable operation, and recycling of the sulfuric acid produced.

ACKNOWLEDGMENTS

This work was funded by STW project 6353 and ERC 232937. Bart Kraakman is thanked for providing samples. Jorge Valdés and David Holmes are thanked for the $A. thiioxidans$ ATCC 19377 and $A. caldus$ ATCC 51756 sequences. Markus Schmid is thanked for advice on primers, and Wendel Brock is thanked for analyzing 16S IR sequences. Jelle Egeneyen is acknowledged for total carbon measurements. We thank Nardy Kip for help with the pH optimum experiment and Geert-Jan Janssen for the cryo-SEM. Sacha van Hijum is thanked for genome assembly, and Daan Speth is thanked for providing an annotated protein list from the draft genome assembly.

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