Union Makes Strength: A Worldwide Collaborative Genetic and Clinical Study to Provide a Comprehensive Survey of RD3 Mutations and Delineate the Associated Phenotype

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Abstract

Leber congenital amaurosis (LCA) is the earliest and most severe retinal degeneration (RD), and the most common cause of incurable blindness diagnosed in children. It is occasionally the presenting symptom of multisystemic ciliopathies which diagnosis will require a specific care of patients. Nineteen LCA genes are currently identified and three of them account for both non-syndromic and syndromic forms of the disease. RD3 (LCA12) was implicated as a LCA gene based on the identification of homozygous truncating mutations in two LCA families despite the screening of large cohorts of patients. Here we provide a comprehensive survey of RD3 mutations and of their clinical expression through the screening of a cohort of 852 patients originating worldwide affected with LCA or early-onset and severe RD. We identified three RD3 mutations in seven unrelated consanguineous LCA families - i.e., a 2 bp deletion and two nonsense mutations – predicted to cause complete loss of function. Five families originating from the Southern Shores of the Mediterranean segregated a similar mutation (c.112C>T, p.R38*) suggesting that this change may have resulted from an ancient founder effect. Considering the low frequency of RD3 carriers, the recurrence risk for LCA in non-consanguineous unions is negligible for both heterozygote and homozygote RD3 individuals. The LCA12 phenotype in our patients is highly similar to those of patients with mutant photoreceptor-specific guanylate cyclase (GUCY2D/LCA1). This observation is consistent with the report of the role of RD3 in trafficking of GUCYs and gives further support to a common mechanism of photoreceptor degeneration in LCA12 and LCA1, i.e., inability to increase cytoplasmic cGMP concentration in outer segments and thus to recover the dark-state. Similar to LCA1, LCA12 patients have no extraocular symptoms despite complete inactivation of both RD3 alleles, supporting the view that extraocular investigations in LCA infants with RD3 mutations should be avoided.


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Introduction

Leber congenital amaurosis (LCA, MIM# 204000) is a group of retinal dystrophies, which represents the most common cause of blindness in childhood (10–18%) [1,2]. This disease is responsible for blindness or profound visual deficiency at birth or in the first months of life. Visual outcome ranges from a severe stationary cone-rod disease with poor visual function (VA ≤ light perception; type I) to a progressive rod-cone dystrophy with measurable visual acuity in the first decade of life (20/200 ≤ VA ≤ 60/200; type II) [3]. Currently, 19 disease-causing genes have been mapped and identified that account for 65–70% of the cases in ethnically mixed populations [1,2]; GUCY2D (LCA1) [4], RPE65 (LCA2) [5], SPATA7 (LCA3) [6], AIPL1 (LCA4) [7], LCA5 (LCA5) [8], RPGRIP1 (LCA6) [9,10], CRX (LCA7) [11,12], CRB1 (LCA8) [13], NBN (LCA9) [14–18], CEP290 (LCA10) [19], IMPDH1 (LCA11) [20], RD3 (LCA12) [21], RDH12 (LCA13) [22,23], LRAT (LCA14) [24], TULP1 (LCA15) [25], KCNJ13 (LCA16) [26], IQCB1 [27,28] and MERTK [29]. Typically, LCA is an autosomal recessive condition. Yet, some rare LCA cases have been ascribed to dominant CRX [11,30–31] and IMPDH1 mutations [20].

The implication of LCA genes in retinal dystrophies of later-onset is not uncommon (GUCY2D, RPE65, CRB1, TULP1, CRX, RPGRIP1, AIPL1) [1,2]. Though less frequently, LCA genes may be involved in severe and sometimes embryonically lethal multisystemic conditions affecting the central nervous system, kidneys, bones and/or the heart in addition to the retina (CEP290, IQCB1) [32–33].

The clinical, genetic and molecular heterogeneity of LCA make elucidating genotype-phenotype correlations a major medical issue to predict the ophthalmologic course of the disease, to allow tailor-made surveillance of extracocular systems and to direct genetic counseling. Significant genotype-phenotype correlations have been reported for most LCA genes; for recent reviews see [1,34] but not RD3 which mutations are amongst the rarest causes of the disease with two homozygous mutations reported in two consanguineous families [21,35].

Here, we report the results of an international study aimed at delineating the clinical and molecular spectrum of RD3 mutations in retinal dystrophies.

Patients and Methods

Patients and Controls

A total of 852 unrelated patients ascertained in -or addressed to-10 Centers in Europe (France, Germany, Italy, the Netherlands and Spain), North-America (The United States and Canada) and China were considered in this study including 574 LCA probands who have not had disease-causing mutations identified in the known LCA genes, 96 probands affected with autosomal recessive early-onset severe retinal dystrophies (EOSRD), 150 with autosomal recessive retinitis pigmentosa (RP) and 32 with other retinal dystrophies. The ethnicity of patients and diagnoses are shown in Table 1. Written consents were obtained from participants or legally authorized representatives at each participating Center. The study was conducted at all sites in strict adherence to the tenets of the Declaration of Helsinki and was approved by the respective National Institutional Review Boards: Including the Comité de Protection des Personnes Ile-de-France II, the McGill University Health Centre Research Institute, the Academic Medical Centre of Amsterdam The Netherlands, the Ethics Committee of the Second University of Naples, the Bioethics Committee of the University of Barcelona, the Ethics Committee of Rotterdam and Nijmegen (MEC-2001-23, MEC-2005-371, and MEC-2010-359), the Baylor College of Medicine Institutional Review Board, the Ethics committee of the Medical Faculty of University of Tubingen and the Zhongshan Ophthalmic Center of Sun Yat-sen University.

Screening of RD3

Samples were collected and screened for RD3 mutations independently. Patients of all cohorts but one were screened by direct sequencing using intronic primers designed to amplify the two RD3 coding exons and intron-exon boundaries. The 313 LCA probands of the University of Iowa Carver College cohort were screened by single strand conformation using as controls 151 normal individuals. PCR, direct sequencing and SSCP conditions, as well as primer sequences are available on request.

A variant was predicted to be pathogenic when it showed familial cosegregation with the disease and absence in control individuals and when it was predicted to be damaging by Align DGVD, Polyphen-2, SIFT, SpliceSiteFinder-like, MaxEntScan,
and the Turkish family share a small 1 Mb-long common
performed on 4/5 families indicate that the two Moroccan families
segregating
Alamut Interpretation Software 2.0.

Results
Linkage disequilibrium analysis
Haplotype analysis was performed using 4/5 families segregating
the c.112C>T mutation of the RD3 gene. Microsatellite markers distributed over a 12.6 Megabases (Mb) interval spanning the
RD3 locus on chromosome 1q22.2-q21 (according to the UCSC Genome Browser GRCh37/hg19 assembly) were used for haplotype analysis. Microsatellite markers included (from telomere to centromere); AFM337YA5 (D1S2782), AFM201Y61 (D1S471), AFM310V61 (D1S491), AFM160YA5 (D1S205), AFM190X65 (D1S414) AFMC01Y65 (D1S2610), AFM-
B342Y61 (D1S2700), AFM203Z66 (D1S425), AFMA127W55 (D1S505), AFMS97X61 (D1S2827), AFMO50X66 (D1S2680).

Amplified fragments were electrophoresed on an automatic
sequencer (ABI 3100, Applied Biosystems, Foster City, CA) and analyzed using the GeneScan Analysis 3.7 and Genotyper
softwares. For each marker, the heterozygote frequency and the
size range of alleles were either available from Genethon Linkage
Map [36]. The size of the alleles carried by the members of families NEM1, NEM2, NEM3 and NEM4 was determined relative to the reference DNA sequence obtained from the Centre d’Etude du Polymorphisme Humain (CEPH).

The degree of linkage disequilibrium and the estimation of the age of the mutation were obtained as described previously [37].

Discussion
Until this report, the implication of RD3 in retinal dystrophies
was based on the following three studies: homozygosity for truncating mutations in two consanguineous LCA families [2/39; 21,35], two naturally occurring models of autosomal recessive
EOGSD, the retinal degeneration 3 (rd3) mouse [21,38] and the rod
dystrophy 2 (rd2) collage [39–40]. The identification in our report of three mutations expected to truncate the protein in seven additional families gives strong support to the role of RD3 in LCA. Yet, the mutation rate obtained in a very large cohort of LCA cases originating worldwide (7/574 LCA cases excluding other LCA genes) and consistent identification of homozygous mutations in consanguineous families [21,35, present study], support the very
low frequency of heterozygous RD3 carriers in the general population.

RD3 encodes an evolutionary conserved basic protein of 195
amino acids that is preferentially expressed in the retina [41]. In
the mouse, it exhibits increasing expression through early
postnatal development. In adulthood, it is highly expressed in
cone and rod outer segments where it co-localizes with retinal-
specific guanylate cyclases, GC1 and GC2, respectively. In the
cones, RD3 has been shown to mediate the export of GC1 from
the RD3 Mutations and the Associated Phenotype
haplotype that differed in the Libanese family (Figure 1).
Haplotype analyses and Bayesian calculations allowed estimating that the founder mutation might have occurred 100–150
generations ago.

In addition to RD3 mutations, we identified a A to T
transversion at position 146 of the cDNA resulting into the
substitution of an asparagine into an isoleucine at position 49 of the protein (c.146A>T; N49I). This variant that changes a basic amino acid into an uncharged residue is predicted to be
deleterious by the SIFT, Polyphen-2 and AlignGCVD programs.
The change was identified neither in 200 control individuals nor in
databases listing SNPs. This change was identified in a heterozy-
gous state in an isolated case born to non-consanguineous parents
(Table 3).

Finally, 13 polymorphic variants were identified, 3/13 of which
were not reported elsewhere (Table 4). All three changes (2/3 non-
conservative) were predicted to be benign by the Alamut Interpretation Software.

Phenotype of patients harboring RD3 mutations
The natural history of the disease and clinical data available of
patients harboring RD3 mutations were reviewed (11 patients in 7
families; mean age = 10.2 yrs; age range 1–31 yrs, Table 2). Patients consistently presented with congenital nystagmus, low
vision, sluggish pupillary reflexes, absence of ocular pursuit since
birth, early-onset and long-lasting dizziness, ocular and extraocular signs of Franceschetti, photosensitive (one exception) and mild to moderate
hyperopia. Visual acuity, when measurable, was reduced to
counting fingers, hand-movements or light perception. Reduced
visual acuity hindered recording color vision and visual field
(Table 2). Fundus consistently showed dull retina with salt and
pepper aspect, thin retinal vessels and an early macular
rerearrangement (maculopathy, Figure 2). Spectral Domain OCT
imaging was available in the Tübingen case. The photoreceptor layer was missing, the inner segment/outter segment border was
diminished and an increased backscatter from the choroid was
observed due to RPE-atrophy. On the other hand, inner retinal
layers seemed to show a rather preserved structure. Fundus autofluorescence (FAF) was severely reduced, a proper image
could not be taken.

NNSPLICE and Human Splicing Finder available under the
Alamut Interpretation Software 2.0.

Results
RD3 mutations, changes of uncertain pathogenicity and polymorphisms
Three novel mutations were detected in seven apparently
unrelated LCA families (Table 2). Two out of the three mutations affect codon c.136–138 (exon 2), which encodes the glutamic acid at position 46. The first one consists of a 2 bp deletion expected to result in the substitution of the glutamic acid into an alanine and the apparition of a premature stop codon, 26 amino-acids downstream (c.137–138delAG; p.E46Afs*83). Sequence analysis indicates that the microdeletion is not expected to alter the splicing
of the messenger RNA (Human Splicing Finder, SpliceSiteFinder-
like, MaxEntScan, NNSplice). The second change consists of a G to T transversion expected to change the glutamic acid at position 46 by a signal of termination of the translation (c.136G>T, p.E46*) and/or to create a cryptic donor-splice site which use
would lead to the production of a 141-aa protein lacking aa 46–99
and thus the C terminus of the predicted coiled coil domain
(Human Splicing Finder score 71.2 vs exon 2 acceptor site 96.7).
The third mutation occurred on the mt112–113 CpG doublet
(c.112C>T). It is expected to change the arginine at position 38 by
a signal of termination of the translation (p.R38*) and/or to create a cryptic donor-splice site which use would result in the production of a 132-aa protein lacking amino acids 38–99 including the
putative coiled-coil domain (Human Splicing Finder, SpliceSite-
Finder-like, MaxEntScan). All three mutations are expected to truncate the RD3 protein. It is thus possible that the mutant
mRNAs may be prone to nonsense-mediated decay. It is interesting to note that the E46 position may be hypermutable
(2 different mutations affecting the codon).

All three mutations segregated with the disease in families and
were neither identified in controls nor reported elsewhere.

The c.112C>T mutation was shared by 5/7 of the families segregating RD3 mutations. Indirect studies at the RD3 locus were
performed on 4/5 families indicate that the two Moroccan families
and the Turkish family share a small 1 Mb-long common
Table 2. RD3 mutations and related retinal phenotype.

<table>
<thead>
<tr>
<th>Position</th>
<th>Mutation type</th>
<th>Mutation</th>
<th>Predicted effect on protein</th>
<th>Country of origin</th>
<th>Light behavior</th>
<th>Refraction</th>
<th>Visual acuity</th>
<th>Fundus Aspect</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exon 2</td>
<td>Nonsense</td>
<td>c.112C&gt;T</td>
<td>p.R38*</td>
<td>Morocco</td>
<td>Photoaversion</td>
<td>−1.50 LE; −1.50 RE (measured without cycloplegic)</td>
<td>NA</td>
<td>Macular rearrangement, With abnormal macular pigmentation with starshape Diffuse salt and pepper aspect, thin vessels</td>
<td>Nystagmus since birth, no ocular pursuit, altered pupillary reflexes</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NEM2/II1F/31</td>
<td>Morocco</td>
<td>Photoaversion</td>
<td>NA</td>
<td>LP LRE</td>
<td>NA</td>
<td></td>
<td>Salt and pepper retina, thin vessels, marked macular atrophy</td>
<td>Nystagmus since birth, altered pupillary reflexes, Keratoconus</td>
<td></td>
</tr>
<tr>
<td>NEM2/II2F/27</td>
<td>Photoaversion</td>
<td>NA</td>
<td>LP LRE</td>
<td>NA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NEM3/II2F/12</td>
<td>Lebanon</td>
<td>Photoaversion</td>
<td>+2 RE; +2.75 LE</td>
<td>LP LE; HM RE</td>
<td>NA</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NEM3/II3F/9(8)</td>
<td>NA</td>
<td>+1 LRE</td>
<td>CF; RE; HM LE</td>
<td>NA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NEM3/II4M/2(1)</td>
<td>NA</td>
<td>NA</td>
<td>HM LRE</td>
<td>NA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NEM4/II1F/22(4)</td>
<td>Turkey</td>
<td>Photoaversion</td>
<td>+4 LRE</td>
<td>LP LRE</td>
<td>NA</td>
<td></td>
<td>Macular rearrangement, thin retina vessels, dull retina with no pigmented migrations</td>
<td>Nystagmus since birth, altered pupillary reflexes</td>
<td></td>
</tr>
<tr>
<td>NEM4/II2F/10(3)</td>
<td>Turkey</td>
<td>Photoaversion</td>
<td>+3 LRE</td>
<td>LP LRE</td>
<td>NA</td>
<td></td>
<td>Macular rearrangement, thin retina vessels, dull retina with no pigmented migrations</td>
<td>Nystagmus since birth, altered pupillary reflexes</td>
<td></td>
</tr>
<tr>
<td>F16338/LCA59-2-94F/17</td>
<td>Turkey</td>
<td>Photoaversion</td>
<td>NA</td>
<td>LP LRE</td>
<td>NA</td>
<td></td>
<td>Macular atrophy, RPE atrophy with salt and pepper aspect, some bone-spicule pigment in the mid periphery</td>
<td>Nystagmus since birth, altered pupillary reflexes</td>
<td></td>
</tr>
<tr>
<td>Exon 2</td>
<td>Deletion</td>
<td>c.137–138delAG</td>
<td>p.E46Afs83*</td>
<td>Algeria</td>
<td>Photoaversion</td>
<td>+4 LRE</td>
<td>LP LRE</td>
<td>Macular rearrangement (4 yrs), thin retina vessels, dull retina with no pigmented migrations</td>
<td>Nystagmus since birth, no ocular pursuit, altered pupillary reflexes</td>
</tr>
<tr>
<td>Exon 2</td>
<td>Nonsense</td>
<td>c.136G&gt;T</td>
<td>p.E46*</td>
<td>Mexico</td>
<td>No photoaversion, No nyctalopia</td>
<td>+1.25 LRE</td>
<td>HM LRE</td>
<td>Macular atrophy, attenuated retinal vessels, abnormal RPE in the mid-periphery, optic disk pallor</td>
<td>Nystagmus since birth, DOSF</td>
</tr>
</tbody>
</table>

LE: left eye; RE: right eye; LP: light perception; HM: hand movements; CF: counting fingers; DOSF: digito-ocular signs of Franceschetti; NA: not available.

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the endoplasmic reticulum to endosomal vesicles with the C terminus of GC1 being required for RD3 binding [21,41–42]. The loss of GC1 and GC2 in the photoreceptors of the rd3 mouse which express a truncated unstable rd3 protein resulting from the p.R107* mutation, is consistent with a role of the protein in the trafficking of GC2 in rods as well. As a consequence, like the double GC1/GC2 knock-out, the complete inactivation of both rd3 alleles is expected to preclude cGMP synthesis in both rod and cone outer segments and to hinder recovery of the dark state after light stimulation [21,42–43]. Consistently, the retina of the animal develops normally but both rod and cone photoreceptors start degenerating by postnatal day 14 - i.e. when eyes open – and the degeneration is almost complete by the age of 2–4 months [21,38,44]. Similarly, the rd3 frameshift mutation (p.Pro139Alafs*) of the rcd2 collie [40] is likely to produce an unstable, non-functional rd3 protein leading to the loss of GC expression and trafficking. The retina of homozygous rcd2 dogs develops almost normally, by the age of 2–2.5 months, the outer segments of both rods and cones are completely lost, with rods degenerating faster than cones [45].

Interestingly, these data which support a common mechanism of photoreceptor degeneration in rd3 and GC double knock-out mice [43] are consistent with the phenotypic overlap between LCA12 and LCA1. Indeed, the review of the natural history and clinical data of our patients and that of the two original RD3 families [21,35] delineate a homogeneous phenotype unambiguously consistent with the diagnosis of congenital and dramatically severe cone-rod dystrophy (LCA type I) originally described in LCA1 patients harboring GUGY2D mutations [3]. RD3 mutations identified thus far are expected to truncate the RD3 protein [21,35, present study]. It is tempting to hypothesize that similar to several other LCA genes, mutations with a milder effect on the protein structure and/or function may be responsible for milder phenotypes; for recent reviews see [1,2]. However, none of the unrelated patients affected by various retinal dystrophies of later-onset and/or severity in our cohort (n = 278) and the other two reported series (n = 907) [21,35,40] exhibited disease-causing RD3 mutations. This finding is consistent with the LCA type I-specific involvement of RD3 mutations. Indeed, of the LCA genes, most genes involved in EOSRD and RP account for LCA type II [1].

Table 3. RD3 Changes of uncertain pathogenicity.

<table>
<thead>
<tr>
<th>Position</th>
<th>Mutation type</th>
<th>Mutation</th>
<th>Predicted effect on protein</th>
<th>Conservation of the mutant aa</th>
<th>Predicted effect (SIFT, Polyphen-2, AlignGCVD)</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exon 2</td>
<td>Missense</td>
<td>c.146A&gt;T</td>
<td>p.N49I</td>
<td>Low</td>
<td>Deleterious</td>
<td>1/200 patient (European descent, single heterozygosity)</td>
</tr>
</tbody>
</table>
Until now, four LCA genes account for congenital cone-rod dystrophy (LCA type I): GUCY2D [5], RPGRIP1 [21], CEP290 [46] and RD3 [present study]. Refraction may represent a distinctive feature in LCA type I since patients with GUCY2D mutations consistently have high hypermetropia (>7) with frequent enophthalmia [3,25] whereas those who harbor RPGRIP1, CEP290 or RD3 mutations present with lower or no hyperopia [25,46, present study]. On the other hand, several population-specific mutations were identified in LCA type I genes. In Mediterranean countries where CEP290 mutations are particularly uncommon [46-47], high frequencies of GUCY2D mutations resulting from founder effects have been reported [25,37,48]. With the present report, RD3 mutations were identified in nine consanguineous families, 6/9 of which originate from Mediterranean countries. Five out of the six families harbored the c.112C>T mutation. Considering, the very low frequency of this mutation in populations with different genetic backgrounds, it is unlikely that it results from the recurrent deamination of the cytosine of the nt112–113 CpG doublet [49]. Besides, indirect studies at the RD3 locus were consistent with the transmission of an ancient founder mutation that occurred in the Mediterranean region. The prevalence of a RD3 mutation in Mediterranean countries allows us to improve the diagnostic flowchart drawn to direct the molecular analysis of LCA type I genes previously reported [1,2]. In LCA type I patients originating from this region with moderate or no hyperopia, RD3 should be screened for mutations first, before GUCY2D, RPGRIP1 and CEP290.

With respect to genetic counseling, our data that support a dramatically low RD3 carrier frequency in the general population indicate that the genetic risk in the case of the union between an affected person harboring RD3 mutations and an individual of the general population with no family history of LCA is negligible. Unions between affected persons with LCA and or between LCA and RP patients are not uncommon. The identification of RD3 mutations in one spouse must prompt the search for RD3 mutations, only when the other spouse is affected with LCA type I. Indeed, if the spouse is affected with LCA type II or autosomal recessive RP of later-onset, the genetic counseling can straight-away be reassuring.

The comprehensive survey of RD3 mutations in large cohorts of patients accurately phenotyped by referent clinicians has major clinical implications. Most LCA genes have demonstrated a preferential or specific retinal expression [1,2]. Consistently, their alterations account for isolated forms of the disease. Yet, increasing numbers of disease-causing genes display greater and more wide-spread patterns of expression including CEP290 which mutations are either responsible for non-syndromic LCA or a range of syndromic forms of the disease; for review see [32]. On the other hand, mutations in the IQCB1 gene at the NPHP5 locus have been shown to cause a form of Senior-Loken syndrome characterized by a congenital retinal dystrophy associated with a renal dysfunction of highly variable age of onset (1st to 5th decade) [27,28,33]. These recent findings support the view that the identification of the disease gene may be a valuable prognostic tool in young individuals affected with LCA. With regard to RD3, it is worth noting that none of the patients reported here whose ages ranged from 2 to 31 had renal failure, neurological symptoms or intellectual disability. Owing to the retina-specific pattern of

### Table 4. RD3 Coding SNPs.

<table>
<thead>
<tr>
<th>Position</th>
<th>Polymorphism type</th>
<th>Change</th>
<th>Predicted effect on protein</th>
<th>Rs number</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exon 2</td>
<td>Non-conservative</td>
<td>c.16T&gt;C</td>
<td>p.W6R</td>
<td>rs35649846</td>
<td>dbSNP</td>
</tr>
<tr>
<td>Exon 2</td>
<td>Non-conservative</td>
<td>c.69G&gt;C</td>
<td>p.E23D</td>
<td>rs34422496</td>
<td>dbSNP</td>
</tr>
<tr>
<td>Exon 2</td>
<td>Non-conservative</td>
<td>c.83C&gt;T</td>
<td>p.T28M</td>
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<td>c.584A&gt;T</td>
<td>p.D195V</td>
<td>-</td>
<td>Friedman et al. 2006; This study</td>
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Novel non-conservative changes were regarded as polymorphisms when they did not segregate with the disease in patients and/or were found in control individuals.

References: 1: http://www.ncbi.nlm.nih.gov/SNP/; 2: Friedman et al. 2006. All Sequence change data are based on the RefSeq cDNA sequence NM_183059.2. doi:10.1371/journal.pone.0051622.t004

doi:10.1371/journal.pone.0051622.t004

**Figure 2. Fundus photographs of the affected person FJC/F (Table 2A).** The proband has poor vision since birth with nystagmus and atrophic lesions in the macular area.

doi:10.1371/journal.pone.0051622.g002
expression of RD3, it is likely that the disease may remain restricted to this tissue. In these patients, axiogenic and expensive extracocular explorations must not be systematically prescribed.

The recovery of the dark state in the retina through GC-mediated increase in cytoplasmic GMP concentration in photoreceptor outer segments is a sine qua non condition to maintain living and functional rod and cone cells. Our knowledge regarding the mechanisms of this complex process - which alteration accounts for ~17% of LCA cases - are incomplete. In particular, most of the molecular players and disease mechanisms of light-driven trafficking between the inner and outer segments of photoreceptors are unknown. Interestingly, although trafficking between the inner and outer segments of most of the molecular players and disease mechanisms of light-driven trafficking between the inner and outer segments of photoreceptors are unknown. Interestingly, although

Finally, it is worth remembering that the success of gene replacement and drug therapy to restore vision in naturally occurring RPE65−/− Briard dogs has paved the way for the development of gene and drug therapies for a broad range of eye disorders and made large dogs the species of choice for these developments [50]. In this context, although RD3 mutations are uncommon causes of LCA, the availability of naturally occurring rd3 animal models, among which a large dog (collie), it is likely that gene and drug therapy protocols will be developed to treat patients with RD3 mutations.

Author Contributions
Conceived and designed the experiments: AIH A. Munnich DZ AB RC. Performed the experiments: IP A. Munnich DZ AB RC. Analyzed the data: AIH A. Munnich DZ AB RC. Contributed reagents/materials/analysis tools: NA CE CL A. Megarbane SB CV BF. Wrote the paper: IP A. Munnich DZ AB RC.

References