Association analysis of genetic and environmental risk factors in the cuticular drusen subtype of age-related macular degeneration

Johannes P.H. van de Ven,1 Dženita Smailhodžić,1 Camiel J.F. Boon,1 Sascha Fauser,2 Joannes M.M. Groenewoud,3 N. Victor Chong,2 Carol B. Hoyng,1 B. Jeroen Klevering,1 Anneke I. den Hollander1,5

1Department of Ophthalmology, Radboud University Nijmegen Medical Center, Nijmegen, the Netherlands; 2Department of Vitreoretinal Surgery, Center for Ophthalmology, University of Cologne, Cologne, Germany; 3Department of Epidemiology, Biostatistics, and HTA, Radboud University Nijmegen Medical Center, Nijmegen, the Netherlands; 4Oxford Eye Hospital, University of Oxford, England, Oxford, UK; 5Department of Human Genetics, Radboud University Nijmegen Medical Center, Nijmegen, the Netherlands

Purpose: To assess the association of gender, cigarette smoking, body-mass index, and nine genetic risk variants with cuticular drusen (CD), a well recognized subtype of age-related macular degeneration (AMD).

Methods: A total of 757 patients with AMD, including 217 patients with CD, and 553 control individuals were interviewed with a questionnaire and underwent an ophthalmic examination. Venous blood samples were obtained for genomic DNA extraction, and genotyping was performed of single nucleotide polymorphisms previously associated with AMD. Odds ratios were calculated for patients with CD, using unaffected control individuals as a reference. Furthermore, odds ratios in patients with CD were compared to those in patients with “non-CD” AMD.

Results: The CD subtype of AMD was significantly associated with current smoking as well as variants in the complement factor H (CFH), age-related maculopathy susceptibility 2 (ARMS2), complement factor B/complement component 2 (CFB/C2), complement component 3 (C3), and apolipoprotein E (APOE) genes. In patients with CD, the association with the CFH Y402H risk allele was significantly higher (p=0.022), whereas the association with current smoking was significantly lower (p<0.001) than in the heterogeneous group of patients with “non-CD” AMD.

Conclusions: The AMD subtype of CD was associated with previously identified genetic AMD risk factors. However, the association with the CFH Y402H risk allele appeared to be stronger, whereas the association with smoking was less pronounced when compared to AMD as a whole. This study suggests a more important role for genetic factors than environmental factors in the development of this well defined subtype of AMD. These findings stress the importance of detailed phenotyping in AMD to identify homogeneous AMD subtypes, which may be associated with different risk factors and disease mechanisms. Such studies will improve the accuracy of predictive models and the effectiveness of preventive and therapeutic options in AMD.

Age-related macular degeneration (AMD) is the most common cause of irreversible and progressive visual loss among the elderly in the Western world [1,2]. The abnormalities of this disorder range from discrete drusen deposits and pigmentary changes in early AMD to geographic atrophy and/or choroidal neovascularization (CNV) in the advanced forms.

AMD is a clear example of a multifactorial disease, and a wide variety of risk factors have been associated with the development and progression of AMD. Advanced age, female gender, cigarette smoking, and a high body-mass index (BMI >30) have been reported as the most consistently reproducible demographic and environmental risk factors in AMD [3-7]. Familial aggregation analyses and twin studies have provided clear evidence of heritability, and more recently strong associations were found with the Y402H (rs1061170) polymorphism in the complement factor H (CFH) gene and with the A69S (rs10490924) polymorphism in the age-related maculopathy susceptibility 2 (ARMS2) gene [8-12]. These two allelic variants contribute to late AMD in more than 80% of cases [13,14]. Other genes that harbor established risk variants for AMD include the complement factor B (CFB), complement component 2 (C2), complement component 3 (C3), complement factor 1 (CF1), and the apolipoprotein E (APOE) genes [15-22].

The fact that AMD is highly heterogeneous in its clinical presentation is well recognized. Nevertheless, most studies reporting on the influence of environmental and genetic risk factors analyzed the AMD phenotype as a whole, without attempting to determine these risk factors in more homogeneous subtypes of the disorder. Based on the clinically observed abnormalities, several subtypes of AMD
may be recognized, including polypoidal choroidal vasculopathy, retinal angiomatous proliferation, and cuticular drusen (CD) [23-29]. The latter, formerly known as basal laminar drusen [30], is characterized by the fundoscopic findings of innumerable, small (25 µm to 75 µm), uniformly sized, round drusen [31]. Most commonly appearing in early adulthood, these drusen are easy visualized with fluorescein angiography (FA). In more advanced stages, the multitude of drusen produce a typical “stars-in-the-sky” appearance in early phases of the angiogram [32]. Researchers have estimated that the CD phenotype comprises approximately 10% of the AMD spectrum [29].

In the present study, we investigated whether the AMD subtype of CD displays different environmental and genetic risk factors than AMD as a whole.

**METHODS**

**Subjects:** In this study, we evaluated a total of 757 unrelated patients with AMD, including 217 patients with CD, and 553 control individuals. All subjects were retrieved from the European Genetic Database (EUGENDA), a multicenter database for clinical and molecular analysis of AMD. In the current study, only Caucasian participants from the Nijmegen (the Netherlands) area participated.

Before being enrolled in EUGENDA, all subjects were interviewed with a questionnaire to document their medical history and lifestyle habits, such as BMI and smoking status. Subjects who reported a kidney disease were excluded from the study to preclude including patients with membranoproliferative glomerulonephritis (MPGN) type II. Pupillary dilatation was achieved with topical 1.0% tropicamide and 2.5% phenylephrine before retinal imaging. Digital non-stereoscopic 30° color fundus photography was performed with a Topcon TRC 50IX (Topcon Corporation, Tokyo, Japan). In addition, patients with AMD received FA and high-resolution Fourier-domain optical coherence tomography (FD-OCT), performed with a combined confocal scanning laser ophthalmoscope/FD-OCT device (SPECTRALIS, Heidelberg Engineering, Heidelberg, Germany).

Color fundus photographs of both eyes of all cases were evaluated by two independent reading center graders according to the standard protocol of the Cologne Image Reading Center and Laboratory (CIRCL). AMD was defined by using international standards as described previously [33]. Individuals of similar age as the AMD cases and who exhibited no signs of AMD in either eye were collected as controls. The CD subtype was defined as a symmetric distributed pattern between both eyes of at least 50 scattered, uniformly-sized, small (25 µm to 75 µm) hyperfluorescent drusen on FA in each eye, of which a minimum of 20 drusen are located outside the Wisconsin age-related maculopathy grading template [34]. After the grading was completed, the AMD cohort was divided into a cohort of patients with the AMD subtype of CD and a group of patients with “non-CD” AMD.

This study was reviewed and approved by the local institutional review boards and adhered to the tenets of the Declaration of Helsinki. Written informed consent was obtained from all individuals before they participated in the study.

**Single nucleotide polymorphism genotyping:** Genomic DNA was isolated from peripheral blood leukocytes using standard techniques and stored at −20 °C. Genotyping of single nucleotide polymorphisms (SNPs) in the CFH (rs1410996), ARMS2 (rs10490924), CFB (rs4151667), C2 (rs9332739), C3 (rs2230199), CFI (rs10033900), and APOE (E2 allele; rs7412 and E4 allele; rs429358) genes in the “non-CD” AMD, CD, and control cohorts were performed as previously described [35]. The CFH variant Y402H (rs1061170) was analyzed with direct sequencing of PCR products using forward primer 5'-TCA TTG TTA TGG TCC TTA TTG TTG GAAC ATG CTA GG-3' and reverse primer 5'-AAA GAC ATG CTA GG-3'. These nine SNPs were selected because they were previously associated with AMD [8-12,15-22]. Fourteen percent of the genotypes were done in duplicate, resulting in a concordance of ≥99.9%.

**Statistics:** Genotype frequencies in the control individuals were tested for Hardy–Weinberg equilibrium. Baseline and clinical characteristics were analyzed with standard descriptive statistics, and differences in gender, smoking status, and BMI were analyzed with a multivariate logistic regression analysis to adjust for the covariates age, gender, BMI, and smoking status where applicable. Subsequently, to study the associations of allele frequencies for AMD-associated SNPs among the “non-CD” AMD cohort, the CD cohort, and the controls, a multivariate logistic regression analysis was performed to adjust for the covariates age, gender, smoking status, and BMI. The differences between the three cohorts are presented as odds ratios (ORs) with 95% confidence intervals (95% CIs).

Data analysis was performed using SPSS software, version 18.0 (SPSS Inc., Chicago, IL). The reported p values are two-sided, and a value of < 0.05 was considered statistically significant.

**RESULTS**

Baseline demographics and risk allele frequencies of the “non-CD” AMD (n=540), CD (n=217), and control (n=553) cohorts are depicted in Table 1 and Table 2. The mean age was 76.7 years (range 55–94; standard deviation [SD] 7.4) in the “non-CD” AMD cohort, 69.3 years (range 50–91; SD 10.4) in the CD cohort, and 73.1 years (range 55–92; SD 6.3) in the controls.
### Table 1. Demographics in “non CD” AMD, CD and control individuals

<table>
<thead>
<tr>
<th></th>
<th>Controls</th>
<th>“non CD” AMD</th>
<th></th>
<th>CD</th>
<th>“CD/no n CD” AMD</th>
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<td></td>
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<td>p-value</td>
<td>OR (95-C.I.)</td>
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<td>p-value</td>
<td>OR (95-C.I.)</td>
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<td>p-value</td>
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<tr>
<td>N</td>
<td>553</td>
<td>540</td>
<td>217</td>
<td></td>
<td>77</td>
<td>69.3 (10.40)</td>
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<td>217</td>
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<td>Mean age (SD)</td>
<td>73.1 (6.25)</td>
<td>76.7 (7.42)</td>
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<td>73.1 (6.25)</td>
<td>76.7 (7.42)</td>
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<td>73.1 (6.25)</td>
<td>76.7 (7.42)</td>
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<td>Gender</td>
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<tr>
<td>Male (%)</td>
<td>242 (43.8%)</td>
<td>210 (38.9%)</td>
<td>77 (35.5%)</td>
<td></td>
<td>0.086*</td>
<td>1.36 (0.96–1.92)*</td>
<td></td>
<td>0.232*</td>
</tr>
<tr>
<td>Female (%)</td>
<td>311 (56.2%)</td>
<td>330 (61.1%)</td>
<td>140 (64.5%)</td>
<td></td>
<td>0.086*</td>
<td>1.36 (0.96–1.92)*</td>
<td></td>
<td>0.232*</td>
</tr>
<tr>
<td>BMI</td>
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<tr>
<td>&lt;25 (%)</td>
<td>242 (43.8%)</td>
<td>224 (41.5%)</td>
<td>95 (43.8%)</td>
<td>Ref.</td>
<td>99 (45.6%)</td>
<td>Ref.</td>
<td></td>
<td>99 (45.6%)</td>
</tr>
<tr>
<td>25–30 (%)</td>
<td>249 (45.0%)</td>
<td>235 (43.5%)</td>
<td>92 (42.4%)</td>
<td>0.169†</td>
<td>1.21 (0.92–1.59)†</td>
<td>0.616†</td>
<td>0.92 (0.65–1.30)†</td>
<td>0.079†</td>
</tr>
<tr>
<td>&gt;30 (%)</td>
<td>62 (11.2%)</td>
<td>81 (15.0%)</td>
<td>30 (13.8%)</td>
<td>0.027†</td>
<td>1.58 (1.05–2.36)†</td>
<td>0.635†</td>
<td>1.13 (0.68–1.89)†</td>
<td>0.079†</td>
</tr>
<tr>
<td>Smoking status</td>
<td></td>
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<tr>
<td>Never (%)</td>
<td>257 (46.5%)</td>
<td>230 (42.6%)</td>
<td>99 (45.6%)</td>
<td>Ref.</td>
<td>99 (45.6%)</td>
<td>Ref.</td>
<td></td>
<td>99 (45.6%)</td>
</tr>
<tr>
<td>Past (%)</td>
<td>272 (49.2%)</td>
<td>230 (42.6%)</td>
<td>96 (44.2%)</td>
<td>0.685‡</td>
<td>1.06 (0.80–1.40)‡</td>
<td>0.977‡</td>
<td>1.01 (0.71–1.43)‡</td>
<td>0.845‡</td>
</tr>
<tr>
<td>Current (%)</td>
<td>24 (4.3%)</td>
<td>80 (14.8%)</td>
<td>22 (10.1%)</td>
<td>2.4x10–11‡</td>
<td>5.97 (3.54–10.09)‡</td>
<td>0.032‡</td>
<td>2.06 (1.07–4.00)‡</td>
<td>2.1x10–4‡</td>
</tr>
</tbody>
</table>

Abbreviations: AMD=age-related macular degeneration; CD=cuticular drusen; OR=odds ratio; CI=confidence interval; BMI=body-mass index; Ref.=reference group. * Adjusted for age, body-mass index and smoking status. † Adjusted for age, gender and smoking status. ‡ Adjusted for age, gender and body-mass index. p-values and ORs printed in boldface indicate significant associations.
### TABLE 2. RISK ALLELE FREQUENCIES IN “NON CD” AMD, CD AND CONTROL INDIVIDUALS

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Controls (%)</th>
<th>“non CD” AMD (%)</th>
<th>p-value</th>
<th>OR (95-C.I.)</th>
<th>CD (%)</th>
<th>p-value</th>
<th>OR (95-C.I.)</th>
<th>CD/&quot;non CD&quot; AMD</th>
<th>p-value</th>
<th>OR (95-C.I.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs10490924/ ARMS2A69S</td>
<td>21.9</td>
<td>42.9</td>
<td>2.0×10^-21</td>
<td>2.94 (2.35-3.67)</td>
<td>38.3</td>
<td>1.7×10^-9</td>
<td>2.25 (1.73-2.93)</td>
<td>0.140</td>
<td>0.83 (0.64-1.06)</td>
<td></td>
</tr>
<tr>
<td>rs1061170/ CFHY402H</td>
<td>38.1</td>
<td>57.7</td>
<td>1.9×10^-14</td>
<td>2.17 (1.78-2.65)</td>
<td>64.1</td>
<td>8.6×10^-16</td>
<td>2.88 (2.23-3.73)</td>
<td>0.022</td>
<td>1.35 (1.04-1.74)</td>
<td></td>
</tr>
<tr>
<td>rs1410996/ CFHY402H</td>
<td>56.9</td>
<td>74.8</td>
<td>9.7×10^-15</td>
<td>2.25 (1.83-2.76)</td>
<td>76.8</td>
<td>3.0×10^-11</td>
<td>2.53 (1.93-3.33)</td>
<td>0.209</td>
<td>1.20 (0.90-1.61)</td>
<td></td>
</tr>
<tr>
<td>rs9332739/C2 E318D</td>
<td>4.7</td>
<td>2.5</td>
<td>0.004</td>
<td>0.47 (0.28-0.79)</td>
<td>1.9</td>
<td>0.016</td>
<td>0.38 (0.17-0.83)</td>
<td>0.744</td>
<td>0.86 (0.34-2.15)</td>
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</tr>
<tr>
<td>rs2230199/ C3 R102G</td>
<td>20.8</td>
<td>27.4</td>
<td>0.001</td>
<td>1.46 (1.18-1.80)</td>
<td>27.9</td>
<td>0.013</td>
<td>1.42 (1.08-1.88)</td>
<td>0.965</td>
<td>1.01 (0.77-1.32)</td>
<td></td>
</tr>
<tr>
<td>rs4151667/ CFB H9L</td>
<td>4.9</td>
<td>2.8</td>
<td>0.006</td>
<td>0.49 (0.29-0.82)</td>
<td>2.3</td>
<td>0.027</td>
<td>0.42 (0.19-0.91)</td>
<td>0.750</td>
<td>0.86 (0.35-2.14)</td>
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<tr>
<td>rs10033900/ CFI</td>
<td>48.2</td>
<td>51.8</td>
<td>0.101</td>
<td>1.17 (0.97-1.42)</td>
<td>51.5</td>
<td>0.232</td>
<td>1.16 (0.91-1.49)</td>
<td>0.809</td>
<td>1.03 (0.79-1.35)</td>
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<tr>
<td>rs7412/ APOE2</td>
<td>8.0</td>
<td>10.7</td>
<td>0.115</td>
<td>1.29 (0.94-1.77)</td>
<td>12.1</td>
<td>0.011</td>
<td>1.65 (1.12-2.42)</td>
<td>0.131</td>
<td>1.37 (0.91-2.05)</td>
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<tr>
<td>rs429358/ APOE4</td>
<td>14.4</td>
<td>10.2</td>
<td>0.058</td>
<td>0.74 (0.54-1.01)</td>
<td>11.1</td>
<td>0.047</td>
<td>0.68 (0.46-1.00)</td>
<td>0.660</td>
<td>0.91 (0.59-1.40)</td>
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</table>

Abbreviations: AMD=age-related macular degeneration; CD=cuticular drusen; OR=odds ratio; CI=confidence interval; ARMS2=age-related maculopathy susceptibility 2; CFH=complement factor H; C2=complement component 2; C3=complement component 3; CFB=complement factor B; CFI=complement factor I; APOE=apolipoprotein E. Missings in genotypes are <15%. Data are adjusted for age, gender, body-mass index and smoking status. Risk allele frequencies of rs10490924, rs1061170, rs1410996, rs9332739, rs2230199, and rs4151667 are significantly associated with “non CD” AMD compared to controls. Risk allele frequencies of rs10490924, rs1061170, rs1410996, rs9332739, rs2230199, rs4151667, rs7412, and rs429358 are significantly associated with CD compared to controls. The risk allele frequency of rs1061170 is significantly higher in CD compared to “non CD” AMD.
Current smoking showed an association with CD (p=0.032; OR: 2.06; 95% CI: 1.07–4.00), and this association was significantly lower (p<0.001; OR: 0.32; 95% CI: 0.17–0.58) compared to the “non-CD” AMD cohort. Female gender showed a trend (p=0.086), and no association with BMI was found for CD.

All genotype frequencies conformed to Hardy–Weinberg equilibrium in the control cohort. The risk allele frequency of the CFH Y402H (rs1061170) variant was 64.1% in the CD cohort, which closely approximates the prevalence reported previously in patients extensively affected with CD [25]. A significantly higher CFH Y402H risk allele frequency was found in the CD cohort when compared with the control cohort (p<0.001; OR: 2.88; 95% CI: 2.23–3.73), and when compared to the “non-CD” AMD cohort (p=0.022; OR: 1.35; 95% CI: 1.04–1.74).

The risk allele frequencies of the ARMS2 (rs10490924), CFH (rs1410996), C3 (rs2230199), and APOE E2 (rs7412) variants were significantly higher in the CD cohort compared to the control cohort, and the protective allele frequencies of the C2 (rs9332739), CFB (rs4151667), and APOE E4 (rs429358) variants were significantly lower in the CD cohort compared to the control cohort. These odds ratios were comparable with the “non-CD” AMD cohort, and no significant differences were observed between the CD and “non-CD” AMD cohort for these SNPs. No association with the CFI (rs10033900) risk allele was found in the CD cohort.

**DISCUSSION**

The clinical spectrum of AMD is broad, and this clinical heterogeneity will influence the results of association studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk factors. Improved phenotyping will increase the power of studies on demographic, environmental, and genetic risk fact
In conclusion, the analysis of a large cohort of the CD subtype of AMD has revealed that genetic risk factors affecting the complement system are especially prevalent in these patients. In addition, the environmental risk factor of smoking appears less influential than in AMD in general. These findings stress the importance of detailed phenotyping in AMD to identify homogeneous AMD subgroups, which may be associated with different risk factors and disease mechanisms. Such studies may improve the accuracy of predictive models and the effectiveness of preventive and therapeutic options in AMD.

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2277