The Origin and Nature of Tightly Clustered BTG1 Deletions in Precursor B-Cell Acute Lymphoblastic Leukemia Support a Model of Multiclonal Evolution

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Abstract

Recurrent submicroscopic deletions in genes affecting key cellular pathways are a hallmark of pediatric acute lymphoblastic leukemia (ALL). To gain more insight into the mechanism underlying these deletions, we have studied the occurrence and nature of abnormalities in one of these genes, the B-cell translocation gene 1 (BTG1), in a large cohort of pediatric ALL cases. BTG1 was found to be exclusively affected by genomic deletions, which were detected in 65 out of 722 B-cell precursor ALL (BCP-ALL) patient samples (9%), but not in 109 T-ALL cases. Eight different deletion sizes were identified, which all clustered at the telomeric site in a hotspot region within the second (and last) exon of the BTG1 gene, resulting in the expression of truncated BTG1 read-through transcripts. The presence of V(D)J recombination signal sequences at both sites of virtually all deletions strongly suggests illegitimate RAG1/RAG2-mediated recombination as the responsible mechanism. Moreover, high levels of histone H3 lysine 4 trimethylation (H3K4me3), which is known to tether the RAG enzyme complex to DNA, were found within the BTG1 gene body in BCP-ALL cells, but not T-ALL cells. BTG1 deletions were rarely found in hyperdiploid BCP-ALLs, but were predominant in other cytogenetic subgroups, including the ETV6-RUNX1 and BCR-ABL1 positive BCP-ALL subgroups. Through sensitive PCR-based screening, we identified multiple additional BTG1 deletions at the subclonal level in BCP-ALL, with equal cytogenetic distribution which, in some cases, grew out into the major clone at relapse. Taken together, our results indicate that BTG1 deletions may act as "drivers" of leukemogenesis in specific BCP-ALL subgroups, in which they can arise in multiple subclones at sites that are prone to aberrant RAG1/RAG2-mediated recombination events. These findings provide further evidence for a complex and multiclonal evolution of ALL.

Introduction

Acute lymphoblastic leukemia (ALL) is the most common form of cancer in children, and can be subdivided in T-lineage leukemia (T-ALL) and B-cell precursor leukemia (BCP-ALL). BCP-ALL makes up about 80% of all pediatric ALL cases, and comprises genetically distinct subtypes. These ALL subtypes are characterized by specific genetic abnormalities, including aneuploidies (hyperdiploidy and hypodiploidy) and chromosomal translocations leading to ETV6-RUNX1, E2A-PBX1, BCR-ABL1, and MLL gene fusions [1]. An additional layer of complexity was recently revealed by the identification of recurrent copy number alterations (CNAs), including single gene deletions [2–4], some of which are strongly associated with disease progression and outcome [5–7]. Despite the high number of CNAs that has been identified in BCP-ALL, the total number of structural genetic lesions in each individual leukemia sample is usually limited to less than 10 per case. A subset of these genetic lesions, predominantly single gene deletions, may act as ‘drivers’ in the development of BCP-ALL. Genes commonly affected by these driver deletions function in key pathways such as cell cycle regulation (CDKN2A, CDKN2B, RB1), lymphoid development (PAX5, EBF1, IKZF1, RAG1/2), and nuclear hormone receptor signaling (NR3C1, TBL1XR1).

One of the newly identified players in BCP-ALL is the B cell translocation gene-1 (BTG1) [4,8]. BTG1 belongs to the BTG/TOB family of anti-proliferative genes (BTG1-BTG4, TOB1 and TOB2) and expression of their gene products is altered in a variety of different cancers [9]. The BTG1 gene has been reported to be affected by deletions in approximately 10% of the BCP-ALL cases [2,3], and to be targeted by nonsense and missense point mutations in diffuse large B-cell lymphoma [10]. BTG1 was shown to directly interact with multiple transcription factors and modulators involved in the regulation of crucial cellular processes including mRNA turnover and histone modification [11–14].
Furthermore, we recently demonstrated that BTG1 regulates the glucocorticoid receptor (GR)-dependent transcriptional response in leukemic cells [8], which turns this gene into an interesting target for modulating therapy response. Several recurrent chromosomal abnormalities in leukemia show a clustering of their breakpoints, suggesting that features intrinsic to the DNA may underlie their origin. A common mechanism underlying these abnormalities is non-homologous end joining (NHEJ). NHEJ is employed by human cells to repair DNA double strand breaks caused by external damage or by intermediate steps of V(D)J recombination and class switch recombination (CSR) events during lymphocyte development [15–17]. The RAG complex mediates V(D)J recombination through the recognition of recombination signal sequences (RSS) located directly adjacent to the V, D, or J segments, thereby generating the variability of immunoglobulins (Ig) as well as B- and T-cell receptors. Illegitimate RAG-mediated recombination has been proposed to be involved in the translocation process of the LM02 and TAL1 proto-oncogenes with TCRβ [18], and appears to be the cause of recurrent intragenic deletions of the IKZF1 gene in BCP-ALL and lymphoid blast crisis in chronic myeloid leukemia (CML) [19,20]. Comprehensive genomic analyses of relapsed BCP-ALL have revealed that diagnosis and relapse samples are clonally related, but may exhibit subtle differences, frequently involving lesions detected at the time of diagnosis that are absent at the time of relapse [5,21,22]. Furthermore, diagnosis and relapse samples may carry alternative lesions affecting genes like e.g. CDKN2A and PAX5, suggesting that these lesions occur repeatedly during clonal evolution of BCP-ALL [5]. These findings are in line with the generally accepted concept that each leukemic outgrowth comprises a heterogeneous population of leukemic cells of which only a subset has the potential to drive progression of the disease, survive therapy, and/or induce relapse development [23]. Recent studies in BCP-ALL have revealed that this multiclonal architecture supports a complex model of clonal evolution [24,25]. During disease progression and after post-treatment relapse, shifts can occur in the dominance of leukemic subclones, most likely triggered by new selective bottlenecks, whereas the subclonal diversity appears to be maintained.

In the present study, we have characterized the origin and nature of BTG1 deletions in a cohort of 831 pediatric ALL cases. We demonstrate that BTG1 deletions are exclusively found in BCP-ALL, both in the predominant clone and in (multiple) minor subclones, and are strongly associated with ETV6-RUNX1 and BCR-ABL1 positive ALL, but rare in hyperdiploid cases. Furthermore, we show that these deletions most likely arise from illegitimate RAG-mediated recombination, are tightly clustered, and virtually all share their telomeric breakpoint within the second exon of BTG1, resulting in the expression of a read-through transcript that gives rise to highly unstable truncated BTG1 protein. These results provide important insight into the mechanism underlying the occurrence of BTG1 deletions in BCP-ALL and support the recently observed complexity of multclonal evolution of ALL.

Results

**BTG1 Is Targeted by Deletions in Specific Subsets of BCP-ALL**

We screened 831 pediatric ALL diagnosis samples for copy number aberrations in BTG1 (Entrez Gene: 694) using multiplex ligation-dependent probe amplification (MLPA). In 722 BCP-ALL samples, we found a total of 65, predominantly monoallelic, deletions (9%), whereas no BTG1 copy number loss was detected in any of the T-ALL samples (n = 109; P = 0.001; Table 1). In addition, MLPA performed on genomic DNA derived from BCP-ALL (n = 10) and T-ALL (n = 5) cell lines revealed BTG1 deletions in the BCP-ALL cell lines REH, SUP-B15, MUTZ5 and 380, while none could be detected in the T-ALL cell lines (see Materials and Methods). Sequencing of the entire gene in 135 BCP-ALL cases and of exon 2, which contains the major body of the open reading frame, in 158 additional BCP-ALL cases and 44 T-ALL cases revealed no mutations. Furthermore, bisulfit sequence analysis showed absence of BTG1 promoter hypermethylation in 20 BCP-ALL and 5 T-ALL cases.

**BTG1 deletions were found to be unevenly distributed between the different cytogenetic subgroups, being enriched in ETV6-RUNX1 (TEL-AML1) and BCR-ABL1 positive cases, and less frequent in hyperdiploid cases (P<0.001, P = 0.003 and P = 0.002, respectively; Table 1). In addition, targeted copy number analysis of recurrently affected genes in ALL revealed that cases with BTG1 deletions more frequently harbored deletions of ETV6, RUNX1, RB1 and EBF1 (P =0.007, P<0.001 and P<0.001 respectively; Table S1). Together, our data indicate that BTG1 is affected by deletions in specific subsets of pediatric BCP-ALL patients.**

**A Deletion Breakpoint Hotspot Maps within the Second Exon of BTG1**

To further define the size and location of the BTG1 deletions, SNP-based genomic profiling was performed on 24 BTG1 deletion-positive BCP-ALL diagnosis samples and the BCP-ALL derived cell lines REH, SUP-B15, and 380. In nearly all cases, the telomeric deletion breakpoint was located within the BTG1 gene. Subsequent PCR-based fine-mapping and direct sequencing revealed that the intragenic breakpoints tightly clustered within a region of 33 base pairs in the second (and last) exon of BTG1, with the majority (75%) of the breakpoints being located within a stretch of 10 base pairs (Figure 1A, Table S2). The centromeric breakpoints mapped to 8 different positions downstream of the BTG1 gene, resulting in deletions ranging in size from 101 to 557 kb (deletions I-VIII). This highly specific clustering of deletion breakpoints allowed us to perform a PCR-based screening of all 65 BCP-ALL diagnosis samples. Hence, we were able to exactly map the deletions in 52 of the 63 cases and found that deletion III was most prevalent, comprising almost half of all lesions identified (49%). Deletions V and VIII were found in 15 and 17% of the...
cases, respectively (Figure 1B). These findings illustrate the high degree of clustering of BTG1 deletion breakpoints in BCP-ALL.

Subclones with Distinct BTG1 Deletions Frequently Arise in BCP-ALL Subtypes

Remarkably, while validating the BTG1 deletions with break-point-spanning PCRs, multiple BTG1 deletions were identified in 18 of the 65 deletion positive cases (27%), all of which turned out to be monoallelic losses using MLPA. The individual deletions appeared to be unique in their exact breakpoints and flanking interstitial sequences (Table S2), thus indicating the presence of multiple subclones which independently acquired BTG1 deletions. In one patient, as many as 4 unique deletions were identified (Figure 2A, Table S3). Triggered by this finding we performed a similar PCR-based BTG1 deletion screening for the three most common deletions (III, V, and VIII) in 89 deletion negative BCP-ALL cases as scored by MLPA or SNP array and were able to confirm the presence BTG1 deletions in minor subclones in 16 cases (18%), which is substantially more frequent than the occurrence of this abnormality in the predominant leukemic clone (Figure 2B). Again, we found unique BTG1 deletions both in the predominant clone and/or in (multiple) subclones (Table 2). An additional deletion was sequenced of which the telomeric break-point is located 2 kb downstream in the 3′UTR of BTG1.

Backtracking and sequencing of the deletion breakpoints in matched diagnosis samples revealed that these BTG1 deletion clones originated from a (sub)clone at diagnosis in at least four cases. These data confirm that BTG1 deletions arise repeatedly and independently during disease progression and that these leukemic (sub)clones frequently reoccur at relapse.

Truncated BTG1 Transcripts Are Expressed in BCP-ALLs with BTG1 Deletions

Virtually all deletions affecting the BTG1 locus target the second exon of BTG1, leaving the possibility of expression of a truncated

Table 1. BTG1 microdeletion occurrence within cytogenetic subgroups of ALL.

<table>
<thead>
<tr>
<th>Cytogenetic subgroup</th>
<th>Number of Samples</th>
<th>BTG1 deletion positive cases (%)</th>
<th>BTG1 deletion negative cases (%)</th>
<th>P-value Chi-square</th>
</tr>
</thead>
<tbody>
<tr>
<td>T-ALL</td>
<td>109</td>
<td>0 (0)</td>
<td>109 (100)</td>
<td>0.001</td>
</tr>
<tr>
<td>BCP-ALL</td>
<td>722</td>
<td>65 (9.0)</td>
<td>657 (91.0)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Hyperdiploid (&gt;50chr)</td>
<td></td>
<td></td>
<td></td>
<td>0.002</td>
</tr>
<tr>
<td>Yes</td>
<td>160</td>
<td>5 (3.1)</td>
<td>155 (96.9)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>477</td>
<td>55 (11.5)</td>
<td>422 (88.5)</td>
<td></td>
</tr>
<tr>
<td>ETV6-RUNX1</td>
<td></td>
<td></td>
<td></td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Yes</td>
<td>142</td>
<td>27 (19.0)</td>
<td>115 (81.0)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>371</td>
<td>22 (5.9)</td>
<td>349 (94.1)</td>
<td></td>
</tr>
<tr>
<td>BCR-ABL1</td>
<td></td>
<td></td>
<td></td>
<td>0.003</td>
</tr>
<tr>
<td>Yes</td>
<td>23</td>
<td>6 (26.1)</td>
<td>17 (73.9)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>625</td>
<td>52 (8.3)</td>
<td>573 (91.7)</td>
<td></td>
</tr>
<tr>
<td>MLL rearranged</td>
<td></td>
<td></td>
<td></td>
<td>0.385*</td>
</tr>
<tr>
<td>Yes</td>
<td>17</td>
<td>0 (0)</td>
<td>17 (100)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>632</td>
<td>55 (8.7)</td>
<td>577 (91.3)</td>
<td></td>
</tr>
<tr>
<td>Other subgroupb</td>
<td></td>
<td></td>
<td></td>
<td>0.155</td>
</tr>
<tr>
<td>Yes</td>
<td>180</td>
<td>13 (7.2)</td>
<td>167 (92.8)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>342</td>
<td>38 (11.1)</td>
<td>304 (88.9)</td>
<td></td>
</tr>
<tr>
<td>Subgroup unknownc</td>
<td></td>
<td></td>
<td></td>
<td>0.245</td>
</tr>
<tr>
<td>Yes</td>
<td>200</td>
<td>14 (7.0)</td>
<td>186 (93.0)</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>522</td>
<td>51 (9.8)</td>
<td>471 (90.2)</td>
<td></td>
</tr>
</tbody>
</table>

BTG1 deletion status was determined using MLPA.

*Because of missing values, numbers do not always add up to 722 BCP-ALL cases. Data was available for 637 cases on hyperdiploidy; 513 cases for ETV6-RUNX1; 648 cases for BCR-ABL1; 649 cases for MLL-rearranged.

†The ‘other’ subgroup encompasses cases negative for ETV6-RUNX1 or BCR-ABL1 translocations, MLL-rearrangement and/or hyperdiploidy. This group includes 10 cases with E2A-PBX1 translocation, of which none harbor a BTG1 deletion.

‡Subgroup unknown includes all cases in which no data is available in one or more cytogenetic classifications.

Fisher’s exact test was used when sample groups were small.

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protein product. To determine whether a truncated BTG1 mRNA was expressed from the rearranged allele, RT-PCR using breakpoint-flanking primers was performed on three BCP-ALL cell lines with a BTG1 deletion (380, REH and SUP-B15 with deletions II, III and IV, respectively) and two cell lines that are BTG1 deletion negative (Nalm6 and RS4;11) (Figure 3). Wild-type BTG1 mRNA was found to be present in all cell lines, whereas truncated read-through transcripts specific for each type of deletion could be detected only in the three deletion positive BCP-ALL cell lines (Figure 3B). Similarly, we detected BTG1 read-through transcripts in primary BCP-ALL samples with BTG1 deletions III, V and VIII, respectively, several of which could be confirmed on genomic DNA of these patients (Figure 3C). Notably, the presence of multiple BTG1 deletion subclones as observed at the genomic level was also detectable at the transcript level and subsequent sequencing confirmed the unique identity of each clone (Table S5). Quantitative RT-PCR revealed that in REH, SUP-B15 and 380 cells the rearranged BTG1 allele was expressed at significantly higher levels compared to the wild-type BTG1 allele, which may reflect differences in mRNA stability (Figure 3D). These results indicate that monoallelic BTG1 deletions result in the expression of truncated BTG1 transcripts, encoding more than half of the BTG1 protein.

To assess the function of the truncated BTG1 protein, a C-terminal deletion variant of BTG1, which terminates at the common breakpoint after amino acid residue 100, was cloned in vector pcDNA3.1 and tested for protein expression. For comparison, wild-type BTG1 was included in the analyses (Figure 1).
Tightly Clustered *BTG1* Deletions in BCP-ALL

A

Number of distinct *BTG1* deletions

<table>
<thead>
<tr>
<th>Percentage of cases</th>
<th>N=1</th>
<th>N=2</th>
<th>N=3</th>
<th>N=4</th>
</tr>
</thead>
<tbody>
<tr>
<td>BTG1 MLPA del (+)</td>
<td>n=34</td>
<td>n=12</td>
<td>6%</td>
<td>3%</td>
</tr>
<tr>
<td>BTG1 MLPA del (-)</td>
<td>n=8</td>
<td>n=5</td>
<td>6%</td>
<td>3%</td>
</tr>
</tbody>
</table>

B

Distribution *BTG1* deletions

- **Predominant clone**
  - Hyperdiploid: 0%
  - ETV6-RUNX1: 19%
  - Total BCP-ALL: 9%
- **Minor clone**
  - Hyperdiploid: 3%
  - ETV6-RUNX1: 50%
  - Total BCP-ALL: 18%
S1A). Full-length HA-tagged BTG1 protein is subject to proteosomal degradation [8], but in the presence of proteosome inhibitor MG132, expression could be readily detected (Figure S1B). In contrast, truncated BTG1 (HA-BTG1-Trunc) protein levels appeared to be highly unstable and hardly detectable even in the presence of MG132 (Figure S1B), while mRNA expression levels were equal to those of full-length HA-BTG1 (Figure S1C). Based on these findings, we conclude that the read-through transcripts that are expressed upon focal loss of BTG1, are unlikely to give rise to functionally active protein, which favors a BTG1 haploinsufficiency scenario.

**BTG1 Deletions May Result from Illegitimate RAG-Mediated Recombination**

Because illegitimate RAG-mediated recombination has been implicated in the origin of several translocations and deletions in leukemia [18–20], we examined the sequences flanking each of the BTG1 breakpoints for the presence of recombination signal sequences (RSS). RSS motifs represent moderately conserved heptamer (CACAGTG) and nonamer (ACAAAAACC) sequences separated by either a spacer of 12±1 bp (12-RSS) or a spacer of 23±1 bp (23-RSS). The RAG complex only joins gene segments containing RSSs with different spacer lengths, complying to the 12/23 rule for efficient V(D)J recombination (Figure S2). Notably, we found that a 23-RSS was present at the recombination hotspot in the second exon of BTG1, while all but one of the centromeric breakpoints were found to harbor a 12-RSS (Table 3; Table S2). Furthermore, all deletion breakpoints contained a random addition of single nucleotides between the joining ends, which is most likely due to the action of terminal deoxynucleotidyl transferase (TdT). Together, these findings strongly support a role for illegitimate RAG-mediated recombination in the occurrence of BTG1 deletions.

The **BTG1 Gene Exhibits Elevated H3K4me3 Levels in B-Lineage Cells**

Recent studies have shown that RAG-mediated V(D)J recombination is controlled by histone modifications, including histone H3 and H4 acetylation and H3 trimethylation, that are normally present at promoter regions of actively transcribed genes [26,27]. More specifically, it has been demonstrated that the RAG2 plant homeodomain (PHD) finger binds directly to H3K4me3, thereby stimulating the catalytic activity of the RAG enzyme complex [28]. To reveal whether differences in levels of H3K4 trimethylation and/or H3K9/14 acetylation could explain the lineage-specific occurrence of BTG1 deletions, chromatin immunoprecipitation (ChIP) experiments were performed targeting different positions within the BTG1 gene in several B-lineage and T-lineage cell lines exhibiting abundant BTG1 expression (Figure 4A). As expected, prominent levels of H3K4me3 were present at the proximal promoter region close to the transcription start site in both B-lineage and T-lineage cell lines (Figure 4B). In contrast, only BCP-ALL cell lines displayed significantly higher levels of H3K4me3 near the breakpoint hotspot within the second exon of BTG1. H3K9/14 acetylation levels were found to significantly differ at the proximal promoter region between B- and T-lineage cells, but not at the body of the BTG1 gene (Figure 4C). In conclusion, our data suggest that illegitimate RAG-mediated recombination at the deletion breakpoint hotspot within the second exon of BTG1 in B-lineage cells may be facilitated by increased levels of H3K4me3.

**Discussion**

In this study, we demonstrate that BTG1, a recurrent target in pediatric ALL, is affected by gene truncating deletions, which occur predominantly in two cytogenetic subgroups of pediatric leukemia.

**Table 2. BTG1 deletions in relapsed cases.**

<table>
<thead>
<tr>
<th>Case</th>
<th>Relapse (MLPA)</th>
<th>Relapse (PCR)</th>
<th>Diagnosis (MLPA)</th>
<th>Diagnosis (PCR)</th>
<th>Backtracking of relapse clone in diagnosis samplea</th>
</tr>
</thead>
<tbody>
<tr>
<td>BCP-ALL 1078</td>
<td>deletion</td>
<td>III, IV</td>
<td>normal</td>
<td>III</td>
<td>two new events</td>
</tr>
<tr>
<td>BCP-ALL 1174</td>
<td>deletion</td>
<td>III</td>
<td>normal</td>
<td>III</td>
<td>match diagnosis and relapse</td>
</tr>
<tr>
<td>BCP-ALL 1193</td>
<td>deletion</td>
<td>IV</td>
<td>normal</td>
<td>N.D.b</td>
<td>one new event</td>
</tr>
<tr>
<td>BCP-ALL 1236</td>
<td>deletion</td>
<td>N.D.a</td>
<td>normal</td>
<td>N.D.a</td>
<td>-</td>
</tr>
<tr>
<td>BCP-ALL 1405</td>
<td>deletion</td>
<td>III, VIII</td>
<td>normal</td>
<td>III, VIII</td>
<td>two new events</td>
</tr>
<tr>
<td>BCP-ALL 1086</td>
<td>deletion</td>
<td>VI</td>
<td>deletion</td>
<td>VI</td>
<td>match diagnosis and relapse</td>
</tr>
<tr>
<td>BCP-ALL 1176</td>
<td>deletion</td>
<td>N.D.a</td>
<td>deletion</td>
<td>N.D.a</td>
<td>-</td>
</tr>
<tr>
<td>BCP-ALL 1304</td>
<td>deletion</td>
<td>III, VIIIb</td>
<td>deletion</td>
<td>VIIIb</td>
<td>new event + match diagnosis and relapse</td>
</tr>
<tr>
<td>BCP-ALL 1273</td>
<td>deletion</td>
<td>N.D.a</td>
<td>deletion</td>
<td>III</td>
<td>new event</td>
</tr>
</tbody>
</table>

*aDetermined by sequence analysis of breakpoint spanning PCR product.

bThe deletion breakpoint could not be detected (N.D.) using the eight breakpoint-spanning PCR assays (I–VIII).

cThe breakpoint does not cluster within BTG1 exon 2, but is located 2 kb downstream in the 3’UTR.

dHomozygous deletion.

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Figure 3. Expression of BTG1 truncated read-through transcripts in BCP-ALL cells with BTG1 deletions. (A) Schematic representation of the wild-type human BTG1 gene, existing of two partly coding exons, and five different BTG1 transcripts due to BTG1 gene deletions. Exons are represented by black (coding) or white (non-coding) bars. Indicated are the RT-PCR primers that were used to detect expression of the wild-type BTG1 transcript (primers A and B), or one of the BTG1 truncated read-through transcripts for deletion II (primers A and C), deletion III (primers A and D), deletion IV (primers A and E), deletion V (primers A and F), or deletion VIII (primers A and G). (B) RT-PCR analyses on total RNA isolated from the BCP-ALL cell lines Nalm6 and RS4;11 (BTG1 wild-type) and REH, SUP-B15 and 380, each with distinct monoallelic BTG1 deletions. (C) RT-PCR analyses on
Tightly Clustered BTG1 Deletions in BCP-ALL

primary BCP-ALL samples in which a single BTG1 deletion (P1, P2, P3 and P4), multiple BTG1 deletions (P4 and P6) or no BTG1 deletions were detected with genomic PCR (P7). Type of deletions (III, V, or VIII) and outcome of MLPA (p: deletion-positive; n: deletion-negative) are indicated. BTG1 read-through transcripts were verified by sequencing (Table S5), except for P3-deletion V, which was an unrelated DNA sequence. (D) Quantitative real-time RT-PCR data representing relative expression levels of BTG1 measured 5’ (primers exon 1/2) and 3’ of the BTG1 breakpoint hotspot (primers exon 2). Expression levels were normalized to Hprt levels, and compared to the expression level in Nalm6, which was set to 1. The data shown represent the average of two independent cDNA reactions and triplicate qRT-PCR reactions.

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Table 3. Authentic RSSs and candidate RSSs flanking breakpoints of BTG1 microdeletions.

<table>
<thead>
<tr>
<th>RSS</th>
<th>Heptamer</th>
<th>Spacer</th>
<th>Nonamer</th>
</tr>
</thead>
<tbody>
<tr>
<td>Optimal consensus</td>
<td>CACAGTG</td>
<td>ACAAAAACC</td>
<td></td>
</tr>
<tr>
<td>VaA27*</td>
<td>CACAGTG</td>
<td>12</td>
<td>ACAAAAACC</td>
</tr>
<tr>
<td>Jκ12</td>
<td>CACAGTG3</td>
<td>23</td>
<td>ACAAAAACC</td>
</tr>
<tr>
<td>TAL2 (9q32)*</td>
<td>CACCTGTG</td>
<td>13</td>
<td>ATAAAAATA</td>
</tr>
<tr>
<td>LM2 (11p13)*</td>
<td>CACAGTA</td>
<td>12</td>
<td>GCATAAAC</td>
</tr>
<tr>
<td>BTG1 exon 2</td>
<td>CACCTCT</td>
<td>23</td>
<td>ACAGAATG</td>
</tr>
<tr>
<td>Deletion I</td>
<td>CACAGTA</td>
<td>12</td>
<td>CCGAGCAT</td>
</tr>
<tr>
<td>Deletion III</td>
<td>CACAGTA</td>
<td>13</td>
<td>ACTGAAATG</td>
</tr>
<tr>
<td>Deletion IV</td>
<td>CACAGGT</td>
<td>12</td>
<td>ACATTTCGA</td>
</tr>
<tr>
<td>Deletion V</td>
<td>CACCTGA</td>
<td>12</td>
<td>CCAATAACC</td>
</tr>
<tr>
<td>Deletion VI</td>
<td>CACAGGTA</td>
<td>13</td>
<td>ACAATAG</td>
</tr>
<tr>
<td>Deletion VII</td>
<td>CACCTGTG</td>
<td>12</td>
<td>ATATATCT</td>
</tr>
<tr>
<td>Deletion VIII</td>
<td>CACAGGT</td>
<td>12</td>
<td>ACAATATG</td>
</tr>
</tbody>
</table>

Mismatches from consensus are underlined; *RSSs flanking VDJ gene segments; *Sequence shown is in reverse complement orientation; *Functional cryptic RSSs at proto-oncogene breakpoints [18].

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prominent deletions, these subclonal lesions displayed a very similar distribution over different cytogenetic subgroups in BCP-ALL and were not present in T-ALL or normal bone marrow samples. Moreover, in one patient, this subclonal BTG1 deletion-positive clone at diagnosis reappeared as the major clone at relapse, whereas in relapse samples of five other patients new (sub)clonal BTG1 deletions appeared. These findings are completely in line with recent studies describing the clonal evolution of ETV6-RUNXI and BCR-ABL1 positive ALLs with a clonal evolution during leukemia development showing that, upon treatment, recurrent lesions occur repeatedly and independently within a single patient, giving rise to a complex variety of slightly different subclones [24,25]. Our current data provide additional evidence for this model at the molecular level by showing that BTG1 deletions arise independently in multiple ALL subclones in a context-dependent manner.

In addition, our results provide insight into the genetic lesions that act in concert with BTG1 deletions in BCP-ALL development. We found, for example, that deletions in ETV6, EBF1 and RBP1 co-occur with BTG1 deletions, indicating that loss of normal BTG1 function may add to defects in pRB/E2F-mediated cell cycle and EBF1-mediated B-cell differentiation pathways. Illegitimate RAG-mediated recombination has been suggested as the responsible mechanism for IZKF1 [19,20], CDKN2A/B [33,34] and LMO2 [35] deletions, as well as ETV6-RUNXI translocations [30]. It remains to be established whether RAG-mediated recombination, or other mechanisms like CSR, are implicated in the occurrence of RBP1 and EBF1 deletions. However, similar to BTG1, these recurrent gene deletions may repeatedly arise in different subclones due to local subtype-specific accessibility of these loci to specific recombination machineries, followed by clonal selection and outgrowth.

In conclusion, our comprehensive analysis of BTG1 aberrations revealed that this gene is recurrently and exclusively affected by deletions in specific BCP-ALL subtypes. BTG1 deletions can arise independently in different ALL subclones, which either develop into a predominant clone at diagnosis, remain present as minor subclones during the course of the disease, or develop into the major clone at relapse. As such, this phenomenon may provide a molecular explanation for the model of multiclone evolution during leukemogenesis.

**Materials and Methods**

**Clinical Samples**

A total of 831 patients diagnosed with BCP-ALL (n = 722) and T-ALL (n = 109) were included in this study. Diagnosis samples (n = 831) and matched relapse samples (n = 62) were collected by the Dutch Childhood Oncology Group (DCOG) and the Radboud University Nijmegen Medical Centre, the Netherlands. Mononuclear cells were harvested through Ficoll gradient separation and DNA was isolated using a QiaAmp purification kit (Qiagen, Venlo, The Netherlands). DNA from non-leukemic bone marrow samples was provided by Dr. Joop Jansen (Department of Hematology, Radboud University Nijmegen Medical Centre, the Netherlands). Written informed consent was obtained for all patient and control samples.

**Cell Lines**

The human BCP-ALL (CCRF-SB, RS4;11, Nalm6, REH, SUP-B15, 380, 697, SEM, TOM1, MUTZ5) and T-ALL cell lines (MOLT4, Jurkat) were purchased either from ATCC or DMSZ. T-ALL cell lines CML-T1, HSB2, and KARPAS45 were obtained from Dr. Adolfo Ferrando (Columbia University, NY, USA) and T-ALL cell line TK6 was obtained from Dr. Albert Fornace (NCI, Bethesda, USA). Leukemia cell lines were maintained in RPMI-1640 medium (Invitrogen) supplemented with 10% fetal calf serum, 100 U/mL penicillin sodium, and 100 µg/mL of streptomycin sulfate at 37°C in a humidified air atmosphere containing 5% carbon dioxide.

**Multiple Ligation-Dependent Probe Amplification**

Leukemic patient samples and cell lines (380, 697, CCRF-SB, RS4;11, SEM, TOM1, REH, MUTZ5, SUP-B15, Nalm6, Jurkat, KARPAS45, MOLT4 and TK6) were analyzed for copy number changes in BTG1 using multiple ligation-dependent probe amplification (MLPA). DNA was isolated using a QiaAmp purification kit (Qiagen). A total of 9 probes was developed using MeltIngenuity software and guidelines provided by MRC-Holland (Amsterdam, The Netherlands; Table S6). MLPA analyses were performed using SALSA MLPA reaction mixture and P200-A1 reference probe-mix (MRC Holland). DNA samples from patients and BCP-ALL samples were used as a positive control. Briefly, 1 µg DNA was incubated with 4 U M.SsI CpG Methyltransferase (New England Biolabs, Beverly, MA) and 250 µM dNTPs at 37°C for 2 h in a 50 µL reaction volume. Reaction mixtures were treated with DpnI methylase to produce an equal amount of unmethylated and methylated DNA. The DNA was then amplified using a set of 19 probes, each with a 10-bp probe sequence at the 5′ end and a 10-bp target sequence at the 3′ end. The probes represent relative expression levels of target genes in the DNA sample. The qPCR reactions were performed in triplicate on an ABI 7500 Fast thermocycler. After initial denaturation at 95°C for 10 min, 40 cycles of denaturation at 94°C for 15 s, annealing at 59°C for 15 s, and extension at 72°C for 30 s were performed. The fluorescence data were analyzed using the ABI 7500 software. The copy number of each target was calculated by comparing the peak areas of the qPCR products to the peak areas of the corresponding standards.

**Bisulfite Sequencing**

In 25 BCP-ALL patient diagnosis samples, we determined the methylation status of the BTG1 promoter using bisulfite sequencing. A total of 500 ng DNA per sample was bisulfite converted using the EZ DNA Methylation™ Kit (Zymo research corporation, Orange, CA, USA). Bisulfite-converted DNA was amplified (for primers see Table S7) and sequenced using an ABI 3730 DNA Sequencer (Applied Biosystems, Foster City, CA, USA). Sequences were analyzed using Vector NTI software (Advanced Genomics Technologies Inc., Foster City, CA, USA).
Ipswich, United Kingdom), 160 μM S-adenosylmethionine, 50 mM NaCl, 10 mM Tris-HCl, 10 mM MgCl₂ and 1 mM Dithiothreitol (pH 7.9) for 4 hours at 37°C. As a negative control, HEK293 DNA was amplified by using the GenomePlex Complete Whole Genome Amplification Kit (Sigma-Aldrich, The Netherlands), according to manufacturer’s protocol.

**BTG1 Mutation Detection**

Sequencing of the first (n = 135) and second (n = 337) exon of the BTG1 gene was performed on PCR-amplified exons using flanking intron-based primers (Table S7). PCR amplifications were performed with 10 ng DNA, 200 nM of each primer, 2 (ex1) or 2.5 (ex2) mM MgCl₂, 200 μM dNTPs, 1 M GC melt (Clontech, Saint-Germain-en-Laye, France), 10× PCR buffer II, 2.5 U AmpliTaq Gold (Applied Biosystems) using 5 minutes preheating at 96°C, followed by 35 cycles of 30 seconds at 96°C, 30 seconds at 55.3°C and 1 min at 72°C, and termination for 5 min at 72°C. The samples were sequenced using a 3730 Sequence Analyzer (Applied Biosystems) and the results were analyzed using Vector NTI software (Invitrogen).

**Genomic SNP Arrays**

To characterize the BTG1 deletions in further detail, we performed high-resolution copy number and genotyping analyses using Affymetrix SNP6.0 arrays according to standard protocols provided by the manufacturer. Genotypes were generated using Affymetrix Genotyping Console v2.1 and Nexus 5.0 software (BioDiscovery, Inc 2010, build version 4621). Normal copy number variation was filtered out using data from our in-house database (containing over 600 healthy control samples), Hapmap, and the Database of Genomic Variants (http://projects.tcag.ca/variation/).

**Breakpoint Mapping and PCR-Based Validation**

Starting from the SNP array data, we used a stepwise approach to zoom in on eight individual breakpoints of different BTG1 deletions. We determined the exact genomic breakpoints and interstitial sequences using Q-PCR, long-range PCR and finally, standard PCR and sequencing. Q-PCR was performed on a 7900HT FAST Real-Time PCR system (Applied Biosystems) with SYBR Green to detect PCR product. Long range PCR was performed using TaKaRa La Taq (Takara Bio Inc, Otsu, Shiga, Japan). PCR amplicons were sequenced using a 3730 Sequencer (Applied Biosystems) and analyzed with Vector NTI software (Invitrogen). The primer pairs used to detect and sequence the eight different breakpoints are listed in Table S7. PCR reactions were performed for 35 cycles at an annealing temperature of 58°C using an Eppendorf Mastercycler. The PCR fragments obtained were resolved by 1% agarose gel electrophoresis, isolated using Qiagen gel extraction kit and cloned into pGEM-T easy for sequencing with M13 primers. Quantitative real-time PCR was performed to determine the expression of BTG1 mRNA sequences 5' to the breakpoint hotspot (exon primers flanking intron 1). BTG1 exon 2 (primers flanking breakpoint hotspot) and HPRT with SYBR Green (Applied Biosystems) (Table S7). Reactions were performed in triplicate on two independent cDNA templates using an Applied Biosystems 7500 Real-Time PCR thermocycler (ABI). Data were represented as fold differences relative to BTG1 mRNA expression in Nalm6 cells normalized to HPRT expression based on calculation of 2^[-ΔΔCt].

**Cloning and Protein Expression Analyses**

**BTG1 Constructs**

HA-BTG1-Trunc was generated by PCR based on full-length HA-BTG1 [8], using primers indicated in Table S7. PCR products were purified, cloned into the EcoRI and Xhol sites of pcDNA3.1 (Invitrogen) and verified by Sanger sequencing. To analyze protein expression of the different BTG1 constructs, HEK293 cells were transfected with 10 μg of plasmid DNA using linear Polyelectrolyteimine (PEI, Polysciences, Warrington, PA). The next day cells were split over two dishes and, 24 hours after transfection, treated with 5 μM MG132 (Peptides International, Louisville, KY) in DMSO or vehicle for 16 hours. For detection of HA-BTG1 protein expression, 5×10⁵ cells were lysed in Laemmli sample buffer containing 2% SDS and 100 mM DTT, boiled for 5 minutes, loaded on 15% SDS-PAGE, blotted on PVDF membrane, and stained with HA antibody, clone 3F10 (Roche Diagnostics). Protein expression was visualized using ECL Plus Western Blot Detection System (GE Healthcare) and FluorchemE digital imaging device (Cell Biosciences).

**Chromatin Immunoprecipitations**

After crosslinking for 15 min with 1% formaldehyde, the leukemia cells were re-suspended in ice-cold lysis buffer (50 mM HEPES [pH7.6], 140 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.1% Sodium Deoxycholate) supplemented with Complete protease inhibitors (Roche) at a density of 35×10⁶ cells/mL and sonicated for 15 min, 30 seconds on and 30 seconds off (Bioruptor Diagenode). Chromatin was incubated overnight in the presence 0.1% BSA, 20 mM HEPES (pH 7.6), 150 mM NaCl, 1 mM EDTA, 0.5 mM EGTA, 0.15% SDS, 1% Triton X-100, protein A/G beads (Santa Cruz), Complete protease inhibitors and 2 μg H3Kime3c or H3K9/14Ac antibody (both from Diagenode). Beads were washed twice in buffer 1 (0.1% SDS, 0.1% Sodium Deoxycholate, 1% Triton X-100, 150 mM NaCl, 1 mM EDTA, 0.5 mM EGTA, 20 mM HEPES [pH7.6]), one time in buffer 2 (0.1% SDS, 0.1% Sodium Deoxycholate, 1% Triton X-100, 500 mM NaCl, 1 mM EDTA, 0.5 mM EGTA, 20 mM HEPES [pH7.6]), one time in buffer 3 (0.5% Sodium Deoxycholate, 0.5% NP40, 250 mM LiCl, 1 mM EDTA, 0.5 mM EGTA, 20 mM HEPES [pH7.6]), and two times in buffer 4 (1 mM EDTA, 0.5 mM EGTA, 20 mM HEPES [pH7.6]). Chromatin was eluted from the beads with 1% SDS/100 mM NaHCO₃ and de-crosslinked together with input material for 4 hrs at 65°C with 200 mM NaCl. After phenolchloroform/isoamylalcohol extraction, DNA was precipitated in the presence of 10 μg glycogen.
Subsequently, diluted DNA was subjected to quantitative PCR using Power SYBR Green (Applied Biosystems) and an Applied Biosystems 7500 Real-Time PCR thermocycler (primers are listed in Table S7). The percentage recovery was equal to dilution factor of input DNA $\times 10^3$. (PDF)

**Statistical Analyses**

Co-occurrence of copy number alterations and cytogenetic subgroups was compared using crosstabs and a standard chi-square test or Fisher’s exact test when sample groups were small. The differences between H3K4me3 and H3K9/14Ac levels in T-ALL versus BCP-ALL samples was assessed using a Student’s t-test. Statistical analyses were carried out using the SPSS statistical package (IBM, Chicago, IL, USA; release 16.0.2, April 2008) and two-sided P-values below 0.05 were considered to be statistically significant.

**Supporting Information**

**Figure S1** BTG1-Trunc protein levels are highly unstable and expressed at extremely low levels. (A) Schematic representation of the full length BTG1 protein and deletion mutant BTG1-Trunc, which mimics the common deletion variant observed in BCP-ALL harboring monoallelic BTG1 deletions. The conserved BoxA, BoxB and BoxC domains as well as the nuclear hormone receptor interaction residues LxxLL are indicated. (B) Immunoblot shows BTG1 protein levels from pcDNA3.1 vectors expressing HA-BTG1-Trunc (Tr) upon transfection in HEK293 cells in the absence or presence of 5 µM MG132 for 16 hrs. Protein expression is detected with HA antibody 3F10. (C) BTG1 RNA expression levels in the same pool of transfected HEK293 cells: Real-time PCR was performed on cDNA generated in the absence (black bars) or presence (grey bars) of Reverse Transcriptase (RT). BTG1 expression was normalized to expression levels of the basal transcription factor TBP. (PDF)

**Figure S2** Monoallelic BTG1 deletions result from aberrant RAG-mediated recombination. The recombination signal sequence (RSS) consists of a conserved 7-bp sequence (heptamer; consensus CACAGTG), a 9-bp sequence (nonamer consensus ACACAAAACC), and an intervening, non-conserved 12 ± 1 or 23 ±1 bp spacer sequence. Recombination that takes place between RSS found in the opposite chromosomal orientation will result in deletion of the intervening DNA sequences. Joining of the RAG-mediated double-strand breaks is carried out by the non-homologous DNA end-joining (NHEJ) proteins via the concerted action of the Ku70/80 proteins, DNA-dependent protein kinase (DNA-PK), Artemis, XRCC4, DNA Ligase 4 (Lig4) and terminal deoxynucleotidyl transferase (TdT). RAG-mediated recombination results in the formation of a precise signal joint and modified coding joint. (A) Schematic representation of the human B cell receptor IgH locus, with the number of gene segments indicated above the V, D and J gene loci. V(D)J recombination occurs only between two gene segments flanked, respectively, by 12-bp RSS and a 23-bp RSS, referred to the 12/23 rule. Thus, the 12/23 rule prohibits direct V(D,J) to-JH joining. (B) Schematic representation of the human BTG1 gene on chromosome 12, which contains a 23-bp RSS at the deletion hotspot in the second exon, and one of the seven identified distal deletion breakpoints harboring a 12-bp RSS. RAG-induced recombination results in deletion of the intervening DNA and a modified coding joint at the BTG1 gene, where TdT adds random, non-templated nucleotides. (PDF)

**Table S1** BTG1 microdeletions co-occur with deletions in recurrently affected genes in BCP-ALL. *Because of missing values, numbers do not always add up to 722 BCP-ALL cases. (PDF)**

**Table S2** BTG1 deletion breakpoint sequences in BTG1 MLPA deletion-positive BCP-ALL cases and cell lines. Sequencing of intragenic BTG1 deletions demonstrates the presence of (near) consensus DNA sequence motifs for V(DJ) recombination flanking the breakpoint hotspot in exon 2 of BTG1 and the distal breakpoint clusters. The consensus heptamer RSS [CAC(A/T)[A/G][C/T][A/G] on (+) strand and [C/T][A/G][C/T][A/G] on (-) strand] is shown in red and bold. Mismatches from consensus are underlined. Single nucleotide mutations are indicated in grey. The nucleotides inserted between the proximal BTG1 and the distal breakpoints are indicated. (PDF)

**Table S3** Number of unique BTG1 deletion-spanning sequences in the BTG1 MLPA deletion-positive BCP-ALL cases and cell lines. (PDF)

**Table S4** Subclonal BTG1 microdeletion occurrence within the cytogenetic subgroups. All patients screened were BTG1 deletion-negative as determined by MLPA. Subclonal BTG1 deletion status was determined using the PCR-based detection of deletions III, V and VIII. *Because of missing values, numbers do not always add up to 89 BCP-ALL cases. Data was available for 77 cases on hyperdiploidy; 41 cases for ETV6-RUNXI; 60 cases for BCR-ABL1; 74 cases for MLL-rearrangement. The subother group encompasses cases negative for ETV6-RUNXI, MLL, BCR-ABL1 translocations and/or hyperdiploidy. This group does not contain any E2A-PBX1 translocation cases. Subgroup unknown includes all cases in which no data is available in one or more cytogenetic subgroups. ^Fisher’s exact test was used when sample groups were small. (PDF)

**Table S5** BTG1 fusion transcript sequences detected in independent subclones. Deletion spanning sequence was confirmed by sequencing of genomic DNA of the same case. (PDF)

**Table S6** MLPA probes for copy-number analysis of the BTG1 gene region. Universal M13 PCR primers are indicated in bold. (PDF)

**Table S7** Primers used for BTG1 bisulfite sequencing, mutation screening, breakpoint mapping, expression analysis and CHIP assay. (PDF)

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**Author Contributions**

Conceived and designed the experiments: EW BS LTvdM PMH AGvK. Performed the experiments: EW BS SVvE LVE YK. Analyzed the data: EW BS FNvL RPK. Contributed reagents/materials/analysis tools: BS ES PMH. Wrote the paper: EW BS RPK.

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References


